



Thesis for the Degree of Master of Fisheries Science

Growth and Survival Rate of *Marphysa sanguinea* (Polychaeta: Eunicidae) Juveniles by Tidal Water Drainage

by

Hortence Koussaye Diatta

KOICA-PKNU International Graduate Program of Fisheries Science

Graduate School of Global Fisheries

Pukyong National University

February 2017

Growth and Survival Rate of *Marphysa* sanguinea (Polychaeta: Eunicidae) Juveniles by Tidal Water Drainage

바위털갯지렁이 치충의 성장과 생존율에 미치는

간출의 영향

Advisor: Prof. Chang-Hoon Kim

by

Hortence Koussaye Diatta

A thesis submitted in partial fulfillment of the requirements for the degree of

Master of Fisheries Science

in KOICA-PKNU International Graduate Program of Fisheries Science, Graduate School of Global Fisheries Pukyong National University

February 2017

Growth and Survival Rate of *Marphysa sanguinea* (Polychaeta: Eunicidae) Juveniles by Tidal Water Drainage

A dissertation

by

Hortence Koussaye Diatta

Approved by:

(Chairman) Prof. Park Wongyu.

(Member) Prof. Kim Jong-Myong.

(Member) Prof. Chang-Hoon Kim.

February 24, 2017

Tables of Contents

Tables of Contentsi
List of Figuresii
List of Tablesiv
Abstract ····································
1. Introduction1
2. Materials and Methods5
2.1 Study Area5
2.2 Juvenile Rock Worms and Experimental design7
2.3 Data collection8
2.4 Statistical Analysis8
3. RESULTS
3.1 Growth performance and Survival rate13
3.2 Abundance of Copepods13
3.3 Water Quality Parameters14
4. Discussion and Conclusion24
References 28
Acknowledgements

List of Figures

Fig. 1. General morphological features and structure of polychaetes2
Fig. 2. Fisheries and science technology centre of Pukyong National University
(PKNU)
Fig. 3. Collection of worms (A) Specimen collection of individual worms
siphoned9
Fig. 4. Experimental set-up (A) : Arrangement of boxes (B) nets used to collect
copepods under the reducers and drums used as main recirculating tanks in
the experiment. (C) Pump. (D) Drum of water with pump. (E) Inside of box,
showing inlet water and aerator, (F) Timer for controlling time exposure. $\cdot\cdot 10$
Fig. 5. Experimental water preparation. (A) Net for collecting copepod (B) The
timer controlling the time of exposure connecting to the pump (C) H9682
Seawater Refractometer for checking the salinity (D) Tank for storing
filtered sea water (E) Tank for storing fresh water (Tap water) (F) Net fitted
under reducers for collecting copepods
Fig. 6. Collection of juvenile rock worms for experiments (A) Scooping sediment
bit by bit from box. (B) Collecting worms by hands12

- Fig. 7. Weight gain of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days.16
- Fig. 8. Specific growth rate of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days.17
- Fig 10. Trend of copepod abundance recorded during rearing of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days19
- Fig. 11. Trend of nitrate (NO₂) concentration recorded in rearing water of M. ...20
- Fig. 12. Trend of ammonia (NH₃) concentration recorded in rearing water of *M*. *sanguinea* exposed to tidal water drainage for 120 days.21
- Fig. 14. Trend of pH concentration recorded in rearing water of M sanguinea ...23

List of Tables

Table 1 Weight gain, Specific growth rate and Survival rate of Marphysasanguinea juveniles exposed to tidal water drainage for 120 days1



Growth and Survival Rate of *Marphysa sanguinea* (Polychaeta: Eunicidae) Juveniles by Tidal Water Drainage

Hortence Koussaye Diatta

KOICA-PKNU International Graduate Program of Fisheries Science

Graduate School of Global Fisheries

Pukyong National University

Abstract

A study was conducted to evaluate the effect of different time exposures by tidal water drainage on growth and survival of *Marphysa sanguinea*, the abundance of copepods and relationship between copepod abundance and survival rate of *M. sanguinea*. Triplicate groups of worms (initial weight $0.19 \pm 0.17g$) were reared under five different time exposures (0hr, 1hr, 2hrs, 3hrs, 4hrs and 5hrs) for 120 days in a semi-recirculating system. Oyster shells (70%, 5 mm in diameter) and sand (30%, 2 mm in diameter) were used together as sediment. After the culture period, there were no significant differences in weight gain (WG) and specific growth rate (SGR) among juvenile *M. sanguinea* reared at 0hr, 1hr, 2hrs, 4hrs and 4hrs was significantly higher than those of the control (0hr) (p>0.05). However, there were no significant differences among the survival rates

of juvenile *M. sanguinea* reared at 0hr, 1hr, 2hrs and 5hrs and also among those of 1hr, 2hrs, 3hrs, 4hrs and 5hrs. The lowest copepod numbers were recorded at 5hr exposure and the lowest number was recorded at 0hr exposure of *M. sanguinea* juveniles to tidal water drainage. The copepod abundance reduced with increasing time exposure, however, at 5hrs time exposure, survival rate reduced. Therefore, it could be concluded that the optimum exposure time to tidal drainage for *M. sanguinea* could be between 3hrs to 4hrs for maximum survival.

Keywords: *Marphysa sanginea*, Time Exposure (tidal water drainage), Growth and Survival rate, Copepods.



1. Introduction

Polychaetes are amongst the most abundant of marine organisms. They are distinguished by linear series of external ringlike segments; the grooves between the segments coincide with internal compartments, often separated by transverse septa, containing serially repeated nervous, muscle and excretory systems. These worms are segmented annelids with a pair of leg-like appendages (parapodia, unique to polychaetes) (Fig. 1). They are known to be good indicators of species richness (Olsgart and Somerfield, 2000) and as bio-indicators of the marine environment (Pocklington and Wells, 1992; Giangrande et al., 2005). Polychaetes are distributed in all oceans of the world and at all depths of the water column. They are found in unique habitats, but are especially abundant in the benthos where they constitute 35-70% of macro invertebrate populations (Díaz-Castañeda and Reis, 2014). However, it is a diverse group with many species and they usually constitute a large proportion of the macro benthos.



Fig. 1. General morphological features and structure of polychaetes.

The rockworm, *Marphysa sanguinea* is a large polychaete (order: Eunicida; family: Eunicidae) that can grow up to a length of 40 cm with a lifespan of approximately 90 days (Prevedelli et al. 2007). Juvenile worms develop two weeks after hatching and take approximately 45 days to reach adulthood. During this time, individuals feed, produce additional segments and structures, and grow to about 27 mm in length. It can be found in a variety of benthic environments, including soft muddy sediments around oyster reefs, sandy or muddy tidal flats, among other fouling organisms on dock pilings and buoys, and inside holes of calcareous rocks (Ruppert and Fox 1988). They are cosmopolitan species: distributed globally at temperate to tropical latitudes (Hutchings and Karageorgopoulos 2003; Prevedelli et al. 2007). *M. sanguinea* is widely distributed in the south coast of England, Eastern Scheldt in south-western Netherlands, and along the

Italian coasts (Prevedelli et al., 2007; Hutchings et al., 2011); along the Sado estuary, particularly within former oyster production areas in Portugal (Garcês and Pereira, 2011) and also in the lagoon of Tunis in Tunisia (Monia et al. 2013).

The demand for *M. sanguinea* has increased rapidly in recent years because they are important nutrient sources for stimulating gonad maturation and spawning of fish and crustaceans in hatcheries for aquaculture purposes (Olive, 1999). They have several advantageous features that may make it ideal for use in environmental studies (Jaramillo et al., 2001). They are also used for culture and commercial purposes to reduce the substrate harvesting disturbance, and the great biogeochemical and benthic community impact (Gambi et al., 1994). However, the production costs of *M. sanguinea* in an intensive aquaculture system should be efficient enough to make profits (Nesto et al., 2012).

The copepods are crustaceans, like shrimps or crabs, with the segmented body covered with a chitinous carapace similar to the pieces of armour. Their size ranges from some tenths of a millimetre to a couple of centimetres, although most commonly they are around one millimetre in length. In general, their body is semitransparent, but in occasions can accumulate pigments or droplets of reserve oil, and then have different colours, from reddish or green, to black or iridescent. They are the most frequent organisms of the zooplankton, can be found at all latitudes and depths, and are considered to be the most abundant metazoans in our planet, even more than insects. As with other organisms, growth and lifespan of marine invertebrates varies with food availability, water temperature, salinity, predator abundance, and other environmental factors. The relationship between environmental conditions, such as time exposure to tidal water drainage, and growth of invertebrates has been well studied. Previous studies have also revealed that inter-specific competition may result in density dependent growth (Jensen and Kamerman; 1992). Therefore, this study was designed to evaluate the effects of tidal water drainage] at different time intervals on growth performance and survival rate of *Marphysa sanguinea* juveniles, the abundance of copepods and the relationship between copepod abundance and survival rate of *Marphysa sanguinea*.



2. Materials and Methods

2.1 Study Area

The study was conducted in Fisheries Science and Technology Centre of Pukyong National University (PKNU) located in Goseong-gun, Gyoungsang Nam-do (Fig. 2). It is adjacent to Geoje, Sacheon, Tongyeong, Masan, and Jinju and linked to the Daejeon-Tongyeong Expressway, National Roads No. 14 and No. 33.





Fig. 2. Fisheries and science technology centre of Pukyong National University (PKNU) located in Goseong-gun, Gyoungsang Nam-do.

2.2 Juvenile Rock Worms and Experimental design

Approximately 360 (initial weight $0.19 \pm 0.17g$) juvenile rock worms were obtained from the Fisheries Science and Technology Centre of the Pukyong National University (PKNU) Goseong-gun, Gyoungsang Nam-do. The worms were collected by scooping the sediment siphoning with a pipette (Fig. 3). 20 individuals per box were collected and put in 28 litre ($45cm \times 25cm \times 10cm$) boxes where they were being raised after hatching. PVC pipes were fitted to the rearing tanks and used as water outlet and overflow. The outlet pipes were covered with nets (1mm mesh size). The containers were filled with treated oyster shells to about 15-16cm deep. The set-up is a semi-recirculating system fitted with reducers, making use of about 450 liters of seawater. The boxes were arranged in 3 rows on 3 floors with 6 boxes on each floor (Fig. 4). Seawater was pumped from the Jaran Bay adjacent to the Research Center, and tap water was the sources of water used in the experiment (Fig. 5). The water was constantly aerated and changed partially (30%) every week and totally on monthly basis throughout the experiment. The flow rate of the water was kept constant at 2L min⁻¹.

The rock worm juveniles were reared under different time exposure times to tidal water drainage; 0hr (control), 1hr, 2hrs, 3hrs, 4hrs, 5hrs in triplicate groups for 3 months. A timer was used to control the different time exposures. Nets were put under all reducers to collect copepods after the time exposure, and counted under the microscope. After the

exposure, the tanks were filled with water again and the worms fed with a commercial shrimp feed at 3% of wet body weight once a day. Salinity and temperature were kept constant at 25 psu and 19 °C respectively. The set-up was kept under a 12 hrs dark and 12hrs of light photo-period. pH, NH₃ (Ammonia), NO₂ (Nitrite) and Alkalinity were measured weekly.

2.3 Data collection

At the end of the experimental period, rock worms in each box were weighed and counted to calculate weight gain (WG), specific growth rate (SGR) and survival rate. Worms were weighed after 30 days, 60 days, and finally after 120 days. During the collection process, boxes were emptied of their sediments and individual worms were collected.

2.4 Statistical Analysis

All data were subjected to one-way analysis variance (ANOVA) using SAS program Version 9.1(SAS Institute, Cary, NC, USA) to test the effects of the different exposure times. When a significant effect was observed, a least significant difference (LSD) test was used to compare the means. Treatment effects were considered significant at confidence level of p>0.05. The mean and SD of the mean was calculated for each treatment.



Fig. 3. Collection of worms (A) Specimen collection of individual worms siphoned out worms by pipette. (B) Juvenile worms collected Sediment from A spread on a tray; (C and D) Juvenile worms collected before measuring weight.



Fig. 4. Experimental set-up (A): Arrangement of boxes (B) nets used to collect copepods under the reducers and drums used as main recirculating tanks in the experiment. (C)Pump. (D) Drum of water with pump. (E) Inside of box, showing inlet water and aerator, (F) Timer for controlling time exposure.



Fig. 5. Experimental water preparation. (A) Net for collecting copepod (B) The timer controlling the time of exposure connecting to the pump (C) H9682 Seawater Refractometer for checking the salinity (D) Tank for storing filtered sea water (E) Tank for storing fresh water (Tap water) (F) Net fitted under reducers for collecting copepods



Fig. 6. Collection of juvenile rock worms for experiments (A) Scooping sediment bit by bit from box. (B) Collecting worms by hands.



 \setminus

3. RESULTS

3.1 Growth performance and Survival rate

Growth performance of juvenile *Marphysa sanguinea* exposed to tidal water drainage is summarised in Table 1 and graphically presented in Fig. 7, 8 and 9. There were no significant differences in weight gain (WG) and specific growth rate (SGR) among juvenile *M. sanguinea* reared at 0hr, 1hr, 2hrs, 4hrs and 5hrs exposure to tidal water drainage. From Fig. 7and 8, the highest WG and SGR was recorded in the third month of the study. Survival rate of juvenile *M. sanguinea* reared at 3hrs and 4hrs was significantly higher than those of the control (0hr). However, there were no significant differences among the survival rates of juvenile *M. sanguinea* reared at 0hr, 1hr, 2hrs and 5hrs and also among those of 1hr, 2hrs, 3hrs, 4hrs and 5hrs.

3.2 Abundance of Copepods

The trend of copepod abundance recorded during the rearing of *M. sanguinea* juveniles exposed to tidal water drainage is shown in Fig. 10. Copepod abundance increased in all groups between day 21 and day 29 of rearing juvenile *M. sanguinea* juveniles. However, the lowest copepod numbers were recorded at 5hr exposure of *M. sanguinea* juveniles to tidal water drainage followed by 4hr, 3hrs, 2hrs, 1hrs and 0hrs in order of increasing numbers.

From the 41^{st} to 48^{th} days, copepod numbers were highest at 1h exposure to tidal water drain, of *M. sanguinea*.

3.3 Water Quality Parameters

Mean water quality measurements during the rearing of *M. sanguinea* juveniles exposed to tidal water drainage are presented in Fig. 11 to 14. There were no significant differences recorded in nitrates (NO_{2}), ammonia (NH_{3}), alkalinity and pH in the rearing water at 0hr, 1hr, 2hrs, 3hrs, 4hrs and 5hrs of *M sanguinea* exposure to tidal drainage over the study period.



R ⁴ (%)
86.66 ^b
90.00 ^{ab}
96.66 ^{ab}
.00.00 ^a
00.00 ^a
90.00 ^{ab}

Table	1	Weight	gain,	Specific	growth	rate	and	Survival	rate	of	Marphysa	
sanguinea juveniles exposed to tidal water drainage for 120 days ¹												

¹Values are means from triplicate groups of worms (n = 3):

²Weight gain (%): [(final weight of worm – initial weight of worm) / initial weight of worm] x 100 ³Specific growth rate (%/day) = [(ln final weight of worm – ln initial weight of worm) / days] x100 ⁴Survival rate (%): (final individuals / initial individuals) x 100



Fig. 7. Weight gain of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days.



Fig. 8. Specific growth rate of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days.



Fig. 9. Survival rate of *Marphysa sanguinea* juveniles exposed to tidal water drainage for120 days.



Fig 10. Trend of copepod abundance recorded during rearing of *Marphysa sanguinea* juveniles exposed to tidal water drainage for 120 days.



Fig. 11. Trend of nitrate (NO_2) concentration recorded in rearing water of M.

sanguinea exposed to tidal water drainage for 120 days.



Fig. 12. Trend of ammonia (NH_3) concentration recorded in rearing water of *M*. *sanguinea* exposed to tidal water drainage for 120 days.



Fig. 13. Trend of alkalinity concentration recorded in rearing water of *M sanguinea* exposed to tidal water drainage for 120 days.



Fig. 14. Trend of pH concentration recorded in rearing water of *M sanguinea* exposed to tidal water drainage for 120 days.

4. Discussion and Conclusion

The results of this study demonstrate that exposure to tidal water drainage do not have significant effects on growth of *Marphysa sanguinea*. This is because no significant differences were found in the weight gain (WG) and specific growth rate (SGR) of juvenile *M. sanguinea* reared at the different time exposures to tidal water drainage. Some worms showed relatively small variations in growth under different environmental conditions, and this may also be attributed to their way of feeding. It is well known that the quality and quantity of available food can affect reproduction and growth in marine invertebrates (Minor and Scheibling 1997; Prevedelli and Vandini, 1997; Prevedelli and Simonini, 2000).

Contrary to growth performance, the time exposure to tidal drainage could affect the survival rate of *M. sanguinea*. The results demonstrate a more positive survival of juvenile *M. sanguinea* reared at 3hrs and 4hrs compared to those of 0hr, 1hr, 2hrs and 5hrs. Low survival rate during the time exposure to tidal water drainage could be attributed to tidal height and sediment type. Other factors such as salinity, light, current speed, exposure to high-energy environments, exposure to siphon-cropping, population density, and genotype have also been noted to influence the growth and survival of marine invertebrates. The interrelationships between many of these variables make it difficult to isolate the most important causal factors (Seed, 1980). Temperature, food

quantity, composition (Rhoads and Young, 1970; Whitlatch, 1980, 1981) and the pressure of predation (Neill, 1975; Bell and Coull 1980; Zobrist and Coull 1992) are among the main factors influencing the growth and survival marine benthos.

Watzin (1983) showed that several groups of macro invertebrate larvae and juvenile were the prey of permanently meiofaunal turbellarians, nematodes and copepods. Copepods have been shown to be facultative predators of polychaete larvae in previous studies (Watzin 1983, Dahms et al., 2004). The abundance of copepods decreased with increasing time exposure during the study period. The negative relations between time exposure and growth have been found in majority of the species studied and predation was another factor which may obscure the relations between environmental conditions and growth of the invertebrates (Goss-Custard, 1977, Zwarts and Drent, 1981; Hulscher, 1982; Sutherland, 1982; Zwarts and Esselink, 1989). It is evident that the density of juvenile polychaetes is significantly affected by the increase in the density of meiofaunal predators (Dauvin, 1990; Fogarty et al., 1991; Coull, 1992). The abundance of different polychaete families (i.e. paranoids and spionids with dominant species, P. caspersi) was significantly affected when the number of predators (copepods) increased. Competition for food between deposit feeding macrofauna and deposit feeding meiofauna could also occur, as well as competition for space since both meiofauna and macrofauna juveniles occupy the uppermost of the sediments (Bell and Coull 1980).

The levels of dissolved ammonia were not significantly affected by the exposure time. In freshwater invertebrates, the main toxic action of nitrate is due to the conversion of oxygen-carrying pigments to forms that are incapable of carrying oxygen. Nitrate toxicity to aquatic animals increases with increasing nitrite concentrations and times exposure. In contrast, nitrite toxicity may decrease with increasing body size, water salinity and environmental adaptation. Freshwater animals appear to be more sensitive to nitrate than marine animals. Prolonged exposure to either high ammonia or nitrite levels can lead to poisoning.

Ammonia is extremely toxic and it can irritate and damage the sensitive tissue with prolonged exposure, which could affect their respiration as well as their ability to expel excess ammonia from the body. Nitrite poisoning is closely related to ammonia poisoning because high levels of one often accompany high levels of the other (Howe and Jeffrey, 2009). In *Penaeus chinensis* Juvenile, nitrite and ammonia concentration has been known to decrease with increasing exposure time (Colt and Armstrong 1981). It has been reported toxic to freshwater organisms at concentrations ranging from 0.53 to 22.8 mg/L. Excess ammonia may accumulate in the organism and cause alteration of metabolism or increases in body pH. At higher levels (>0.1 mg/liter NH₃) even relatively short exposures can lead to skin, eye, and gills damage. Slightly elevated ammonia levels falling within the acceptable range may adversely impact aquatic life (Brian, 2014).

In summary, the present study demonstrates that time exposure to tidal drainage does not affect the growth of *M. sanguinea* indicating the ability of the polychaetes community to respond to changes in their environment. However, the time exposure affects their survival and the density of copepods in the rearing environment. The copepod abundance reduced with increasing time exposure, however, at 5hrs time exposure, survival rate reduced. Therefore, it could be concluded that the optimum time exposure to tidal drainage from the results of this study for *M. sanguinea* could be between 3hrs to 4hrs for maximum survival.



References

Brian, O., 2014. Water Research Watershed Center. http://www.water-research.net.

- Bell, S.S and Coull BC., 1980. Experimental evidence for a model of juvenile macrofauna-meiofauna interactions. In: Tenore K.E and Coull, B.C., (eds) Marine Benthic Dynamics. University of South Carolina Press, Columbia S.C, 179-192.
- Colt, J.E and Armstrong, D.A.., 1981. Nitrogen toxicity to crustaceans, fish and molluscs. In: Allen, J.L and Kinney, E.C., (eds.) Proceedings of the Bio-Engineering Symposium for Fish Culture. Fish Culture Section, American Fisheries Society, Northeast Society of Conservation Engineers, Bethesda, Maryland, 34-47.
- Dauvin, J.C., 1990. Recrutement meiobenthique des principales espsces de Polychetes et de Mollusques Bivalves d'un peuplement subtidal de sediments fins de la Manche occidentale. Cahier Biologique Marine 31, 201-224.
- Dahms H.U., Harder T and Qian P.Y., 2004. Effect of meiofauna on macrofauna recruitment: settlement inhibition of the polychaete *Hydroides elegans* by the *harpacticoid copepod Tisbe japonica*. Journal Experimental Marine Biology Ecology 311, 47–61

- Gambi, M.C., Castelli., Giangrande A.A., Lanera, P.D., Prevedelli and Vandini, R.Z.,
 1994. Polychaetes of commercial and applied interest in Italy: an overview. In:
 Actes de la 4ème Conférence Internationale des Polychètes.Memoires du Muséum
 National d'Histoire Naturelle.
- Garcês, J.P and Pereira, J., 2011. Effect of salinity on survival and growth of *Marphysa sanguinea*, Montagu, 1813 juveniles. Aquaculture International 19, 523-530.
- Howe, Jeffrey C. "Aquarium Nitrate Trouble." Fish Channel.com. http://www.fishchannel.om/fish-magazines/freshwater-and-marine aquarium/august 2009/aquarium-nitrate-trouble.aspx.
- Jaramillo, C.A., and D.L. Dilcher, 2001. Middle Paleogene palynology of central Colombia, South America: A study of pollen and spores from tropical latitudes, Paleontographica 258, 87-213.
- Jensen, K.T., 1992. Dynamics and growth of the cockle, *Cerastoderma edule*, on an intertidal mud-flat in the Danish Wadden Sea: effects of submersion time and density. Nethal Journal Sea Response 28, 335-345.
- Kamermans, P., 1992. Growth limitation in intertidal bivalves of the Dutch Wadden Sea. Thesis, University Groningen 1-135.

- Monia, E.B. Patrick, S and Fathia , Z., 2013. Reproductive Cycle of *Marphysa Sanguinea*, Montagu, 1915; (Polycheata: Eunicidea) in the Lagoon of Tunis. The Scientific World Journal 7.
- Nesto, N.R. Simonini, D., Prevedelli, and Daros, L.; 2012. Effects of diet and density on growth, survival and gametogenesis of *Hediste diversicolor*, Müller, O.F., 1776 (Nereididae, Polychaeta). Journal Aquaculture 363, 1-9.
- Olive, P.J.W., 1999. Polychaete aquaculture and polychaete science: a mutual synergism. Hydrobiologia 402, 175-183.
- Olsgart, F and Somerfield P.J., 2000. Surrogates in benthic investigations. Which taxonomic units? J Aquatic Ecosystem Stress Recover 7, 25-42.
- Prevedelli, D., Gloria, M.N., Ivano, A. and Simonini, R., 2007. Life cycle of Marphysa sanguinea (Polychaeta: Eunicidae) in the Venice Lagoon (Italy). Marine Ecology 28, 384 393.
- Rupper, E.E and Fox, R.S.; 1988. Seashore Animals of the Southeast; A guide to Common Shallow-Water Invertebrates of the Southeastern Atlantic Coast. University of South Carolina Press, Columbia 429.

- Seed, R., 1980. Shell growth and form in the bivalvia. In: D.C. Rhoads and R.A. Lutz. Skeletal growth of aquatic organisms. Plenum Press, New York 23-67.
- Watzin M.C., 1983. The effects of meiofauna on settling macrofauna: meiofauna may structure macrofaunal communities. Oecologia 59, 163–166
- Whitlatch, R.B., 1980. Patterns of resource utilization and coexistence in marine intertidal deposit feeding communities. Journal Marine Response 38, 743-765.
- Whitlatch, R.B., 1981. Animal-sediment relationships in intertidal marine benthic habitats: some determinants of deposit-feeding species diversity. Journal Experimental Marine Biology Ecology 53, 31-45.
- Zwarts, L and Drent R.H., 1981. Prey depletion and the regulation of predator density:oystercatchers (Haematopus ostralegus) feeding on mussels (Mytilus edulis). In: Jones N.V. and Wollf W.J. Feeding and survival strategies of estuarine organisms. Plenum Press, New York 193-216.

Acknowledgements

Thanks to KOICA- PKNU Program for the scholarship and funding of this study. I thank my supervisor, Prof. Chang-Hoon Kim for his guidance and supervision during my study and preparation of this thesis. Thanks to my Thesis Advisory Committee – Prof. Jong Myoung Kim and Prof. Wongyu Park for their inputs to the preparation of this thesis. I am very grateful to all my laboratory mates in Busan and Goseong, especially, to Thi Thu Em Vo and Kyeon Hun Kim for their support and assistance in designing and carrying out this experiment. I thank all ones who spent their quality time to read my manuscript.

