

工學碩士 學位論文

Chitin Chitosan

2002 年 2 月

釜慶大學校 大學院

食品 工學科

宋 浩 銖

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指導教授 李 根 泰

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Studies on the Application of Near Infrared Spectroscopy for Analysis of Chitin and Chitosan

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Abstract

Since near-infrared(NIR) spectroscopic analysis, one of the most popular method for nondestructive measurement among various equipment analyses, does not require such treatment processes as pretreatment of using a reagent, it has been actively used in various fields for the development of analysis methods.

Chitin and chitosan, which are recognized as a representative case for the use of by-product of marine process, require deproteinization process and deacetylation process, and the current methods used for the quality management of this process require expenditure of materials or

time. In this research, NIR spectroscopic analysis has been studied for the measurement of deproteinization and deacetylation to apply the merits of NIR spectroscopic analysis to the quality management in the process of chitin and chitosan production, and the results derived from this study are summarized as follows;

First, in measuring squid pen and red snow crab shell, which are raw materials of chitin and chitosan by NIR, there were the typical peaks in 1200nm, 1510nm, 2050nm and 2180nm. Squid pen had somewhat higher peak than red snow crab shell, and it shows the same result as the result of chemical protein analysis.

In producing chitin, the amount of protein was decreased. Measuring it by NIR, reduction of protein caused by deproteinization was identified in producing chitin.

Chitosan is a derivative material made from chitin by processing the deacetylation. During this processing, acetyl groups were removed and amide bonds were appeared. From NIR spectra, peaks at 1530nm and 2030nm indicated amide II peak of chitosan, and these peaks were used for identifying the differences of structure between chitin and chitosan.

Currently, IR is used for the measurement of the deacetylation degree of chitosan. But, the measurement of nonidentified sample by using the standard curve which has prepared by different deacetylation degrees found out the possibility of measurement of the deacetylation degree by

NIR. The error in measurement of nonidentified sample was below 1% and the error in the standard curve was below 0.006. These errors were very low and the accuracy of NIR was considered to be superior to the existing methods.

In using NIR to examine the structural changes by fat absorption of chitosan, it was identified that fat absorption occurred by the combination of C-H group in fat and amide group in chitosan.

가

가

가

가

가

가

가

가

가

가

Dull(1978) “

가 ”

가

,

(Near-Infrared)

가

가 C-H, N-H, O-H

가 $0.8\mu\text{m}$ - $2.5\mu\text{m}$

가

(Lanza.

1983).

1960 Norris Ben-Gera

spectrophotometer

, Near Infrared Reflectance Spectroscopy (NIR)

가 .

10

가

(NIR)

.

,

가

.

NIR

5

가

가

가

가

NIR

가 가

가

가

가

chitin

chitosan

cellulose

가

가

. Chitin

가

chitin

chitosan

가

(Table 1).

chitin

chitosan

Table 1. Industrial applications of chitosan.

Application fields	Available functions
Wastewater Treatment	Removal of metal Flocculant and coagulant <ul style="list-style-type: none">- protein- dye- amino acids
Food Industry	Removal of dye and suspended solids etc Preservative Color stabilization Animal feed additive
Medical	Bandages Blood cholesterol control Controlled release of drugs Skin bum Contact lens
Biotechnology	Enzyme and cell immobilization Protein separation Cell recovery Chromatography
Agriculture	Seed coating Fertilizer Controlled agrochemical relaese
Cosmetic	Moisturizer Face, hand and body creams Bath lotion Hair care
Pulp and Paper	Surface treatment Photographic paper
Membrane	Permeability control Reverse osmosis

,
 .
 chitin chitosan
 chitin chitosan (Fig.1), chitosan
 , 가
 chitin chitosan
 .

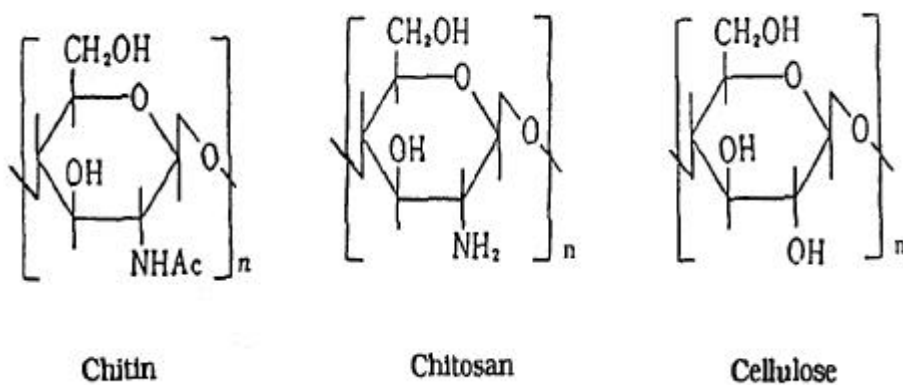


Fig. 1. Chemical structure of chitin, chitosan and cellulose.

I.

1- 1.

가 (Ommastrephes bartramii) 40 12
50mesh chitin .

1- 2.

Chitin chitosan 가 가
(Chironomocetes japonicus)
50 , 20mesh
(5) .

.

1. chitin

50mesh 1N NaOH (15) 가
3
40 chitin (Fig.2).

2. chitosan

Chitin 100g 40% NaOH 1.6 가 110 115 3
chitosan (Fig.2).

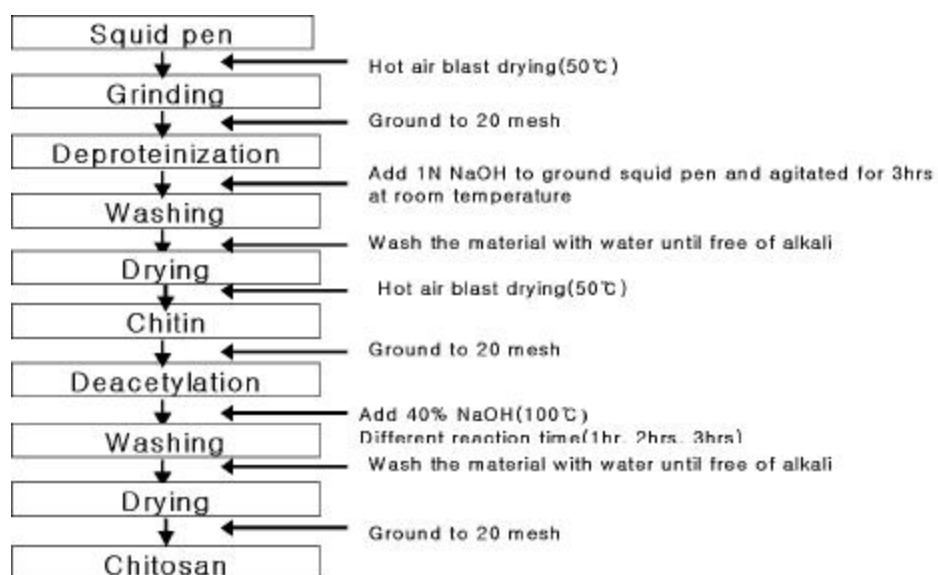


Fig. 2. Preparation procedure of chitin and chitosan from squid pen.

3. chitin

20mesh . 2N HCl
10 가 3 1N NaOH
100 3 pH가
50 chitin (Fig.3).

4. chitosan

Chitin 20mesh chitin 47% NaOH 13
0 가 가 1 , 2 , 3
50
(Fig.3).

5. chitosan

chitosan 0.1M acetate - 0.1M sodium acetate
buffer(pH 4.0) 1% lipase chitosan 1
: 0.3 40 24 incuvation ,
100 15 autoclaving . Inactivation 가
glass filter chitosan , chitosan
0.1M NaOH 가 (5000rpm, 10)
(Vacuum Freezer
dryer, SFDSM12, Samwon Freezing Engineering Co.)
(Fig.4).

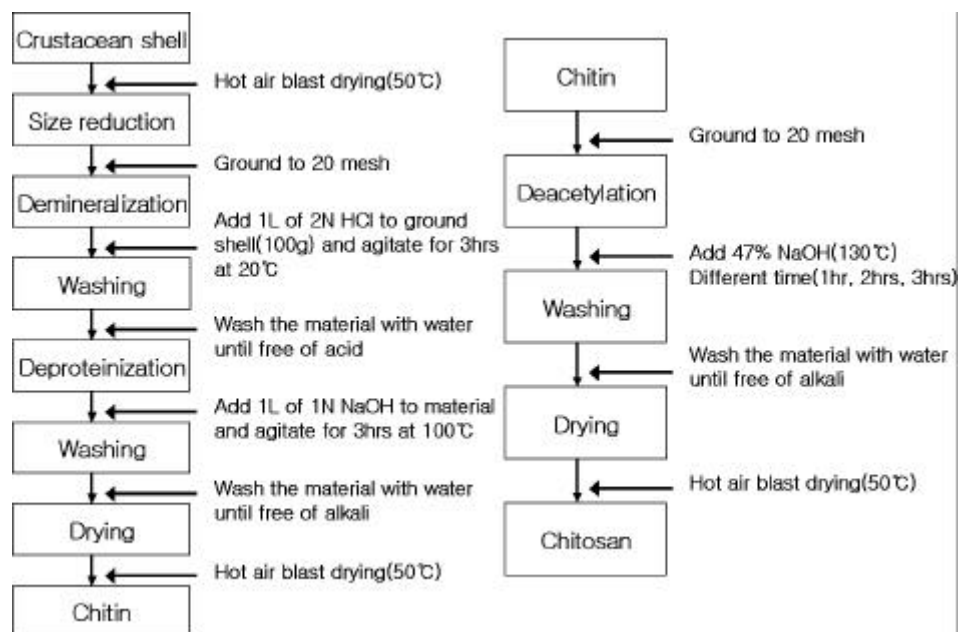


Fig. 3. Preparation procedure of chitin and chitosan from red snow crab shell.

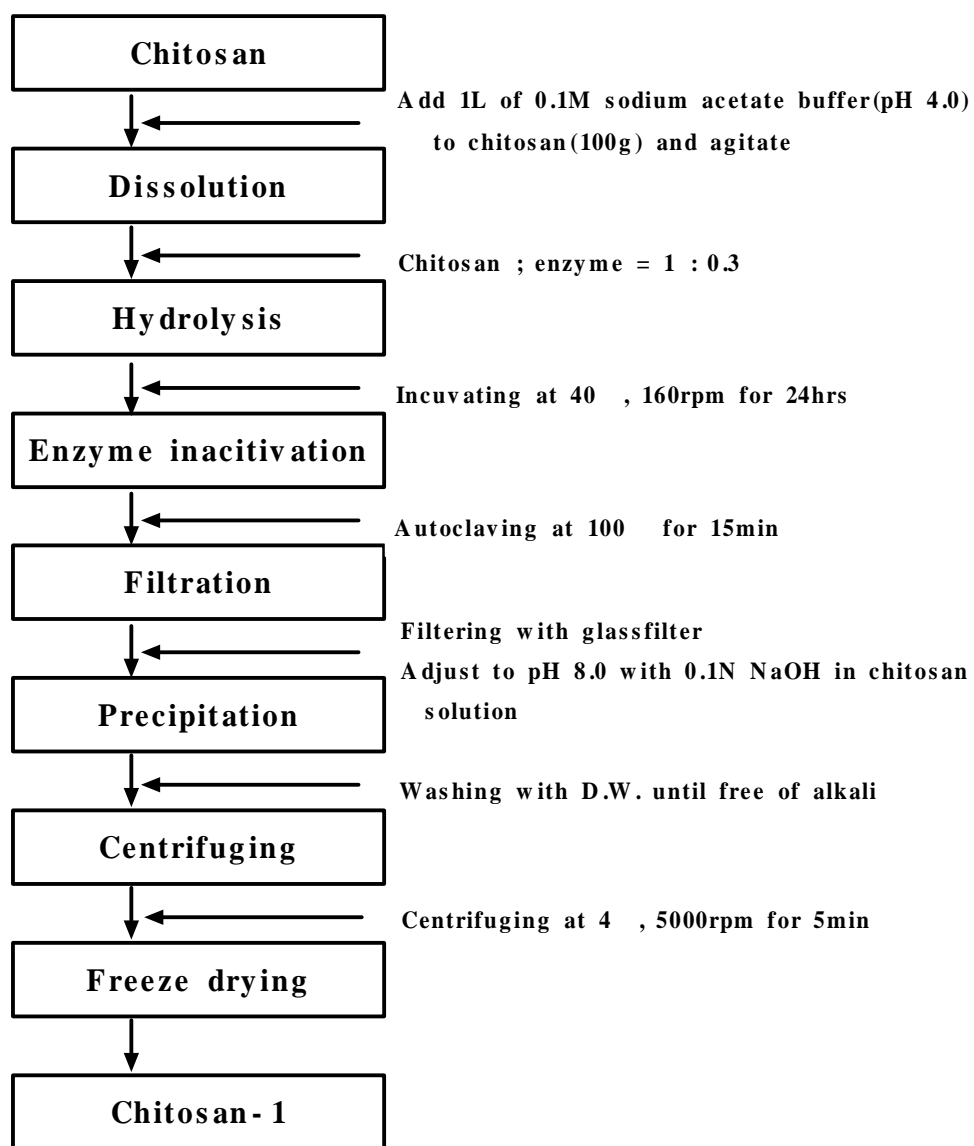


Fig. 4. Preparation procedure of low molecular weight chitosan.

6.

가. (Moisture)

AOAC(1990) dry oven 105 가

. (Crude protein)

AOAC(1990) semi-micro Kjeldahl

chitin chitin .

. (Crude fat)

AOAC(1990) Soxhlet

. (Crude ash)

AOAC(1990) .

7. Chitin chitosan IR spectrum

chitin	chitosan	chitin	chitosan
200mesh	KBr	(200mesh) 100mg	standard pellet
	KBr	100mg	2mg vibration mill
10		pellet	FT - IR (Spectrum GX, Perkin
Elmer, USA)	4,000cm ⁻¹ 500cm ⁻¹		(Fig 5).
Sannan	(1978)	IR spectrum	2,878cm ⁻¹

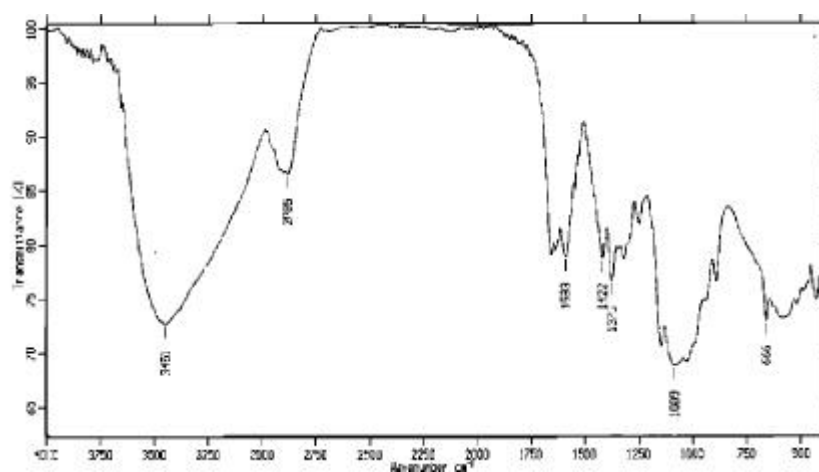


Fig. 5. Typical infrared spectra of chitosan.

1,550cm⁻¹ (A₁₅₅₀/A₂₈₇₈)

(Fig.6).

8. Chitin chitosan

Chitin chitosan chitosan
powder
Standard sample cup (Spectrum GX,
Perkin Elmer, USA) . 1100 - 2400nm
absorbance mode 2nm 64 scan

9. chitosan

Lin (1974) 0.1M acetate-0.1M sodium acetate buffer(pH 4.0)
500Mℓ chitosan 5g 50g () 가
1N NaOH pH 7.0 .
(3000rpm, 25min) 45 ° 30
(SFDSM12, Samwon Freezing Engineering Co.)
pellet .

10.

NIR (estimated value) (actual value)
PCR PLUS(Perkin Elmer, USA)

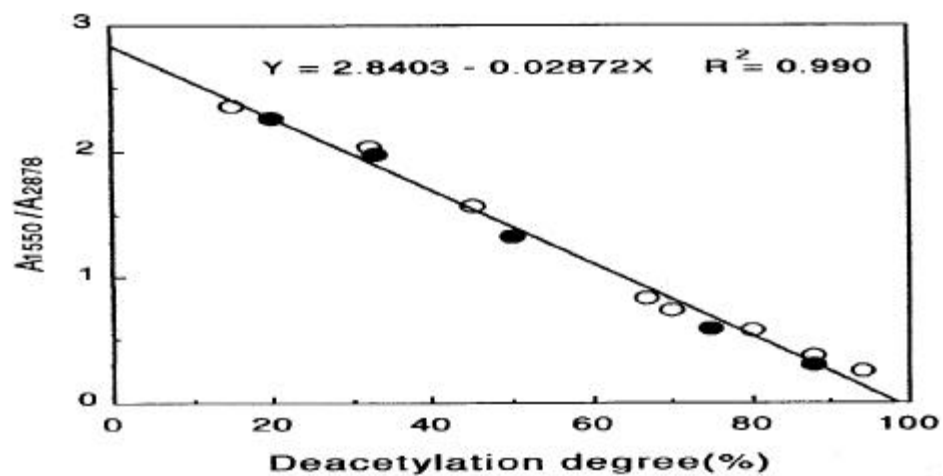


Fig. 6. Calibration line obtained by plotting the ratio of the absorbance of the bend at 1550cm^{-1} to that at 2878cm^{-1} against the degree of deacetylation (from Sannan et al., 1978).

1.

Chitin

가

chitin

Table 2

Table 3

46.2%

0.8%

chitin

12.1%,

8.3%

1100nm 2500nm

2nm

64 scan

Fig.7

1500nm, 2180nm

peak가

peak

N-H

peak가

. Robertson (1989)

1200nm, 1510nm, 2050nm, 2180nm

peak가

2. Chitin

chitosan

2-1

chitin

chitin

Fig.8

chitin

Table 2. Ingredients and yield of dried shell and chitin from red snow crab (dry basis)

Composition	Shell	Chitin
Yield(%)	100	22.3
Moisture (%)	9.4	8.1
Nitrogen (%)	8.3	6.76
Crude ash (%)	46.2	0.3

Table 3. Ingredients and yield of squid pen and chitin from squid pen (dry basis)

Composition	Squid pen	Chitin
Yield(%)	100	36.1
Moisture (%)	5.6	3.7
Nitrogen (%)	12.1	8.1
Crude ash (%)	0.8	0.4

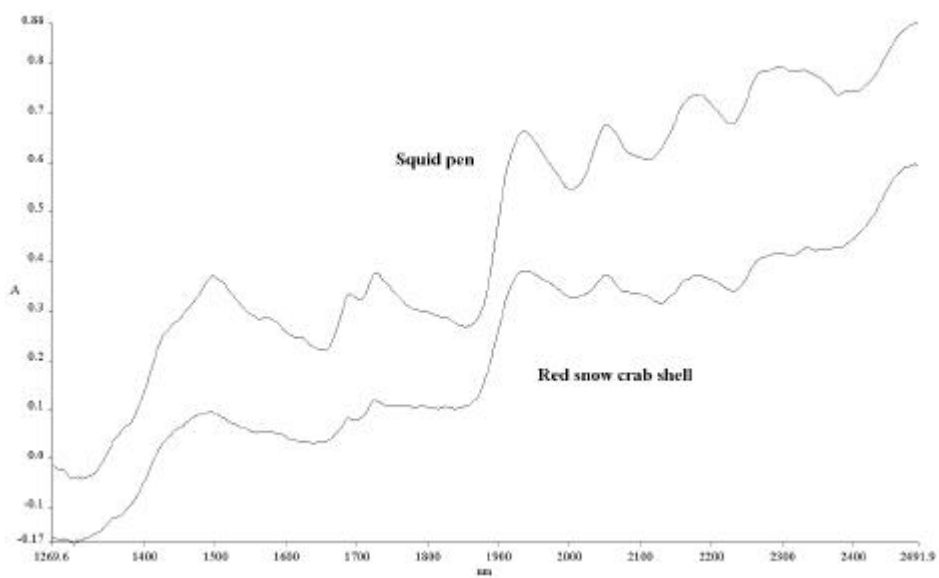


Fig. 7. Near-infrared spectra of red snow crab shell and squid pen.

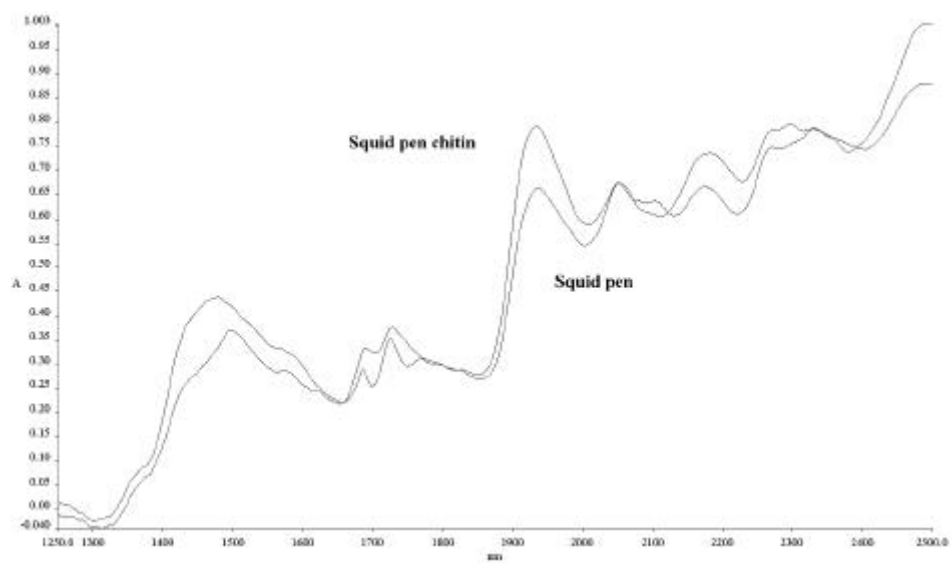


Fig. 8. Near-infrared spectra of squid pen and chitin.

1500nm 2180nm 가 .
 N - H peak peak가
 chitin . chitin
 chitin
 chitin
 N - H peak가
 .
 1930nm chitin peak가
 O - H
 chitin O - H 가
 chitin
 chitin

2 - 2

chitin chitosan

chitin chitosan spectrum
 Fig.9 . chitosan 1530nm 2030nm
 amide peak가 .
 Chitin chitosan 2 Fig.10
 . peak
 가 . 2
 chitin chitosan peak 1530nm 2030nm
 amide peak가 .
 chitosan

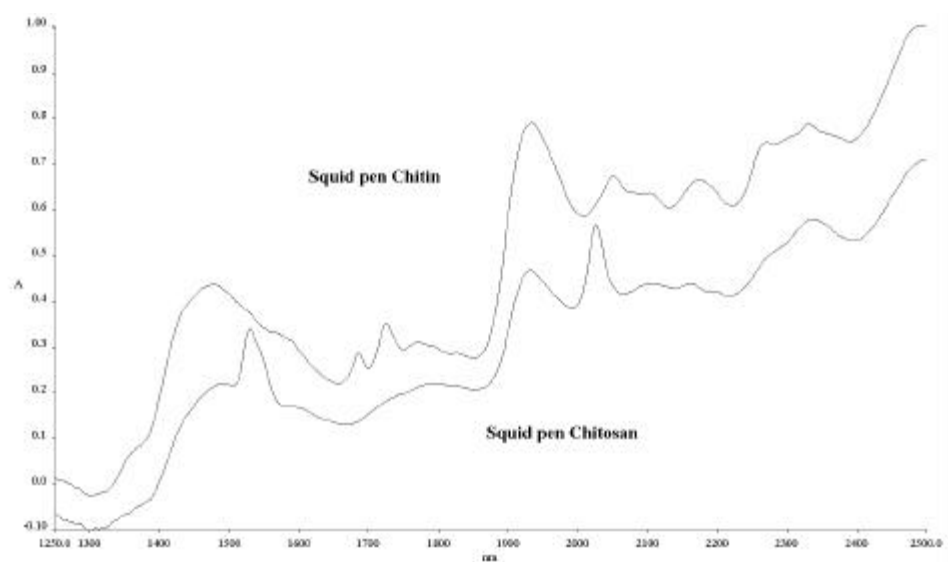


Fig. 9. Near-infrared spectra of squid pen chitin and squid pen chitosan.

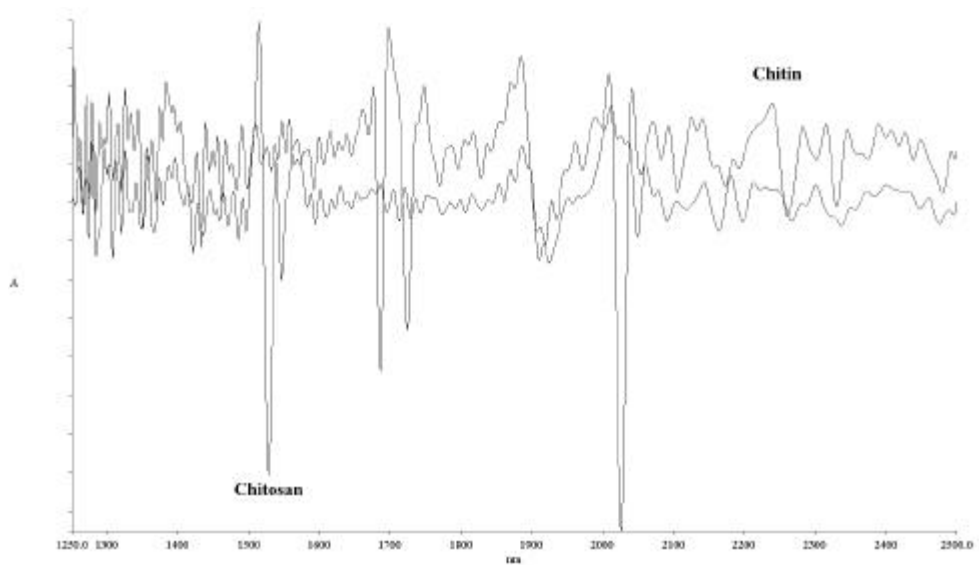


Fig. 10. Second derivative near-infrared spectra of squid pen chitin and chitosan.

chitosan spectrum (Fig.11), chitosan 1530nm 2030nm amide peak가 . chitosan , chitosan 1530nm 2030nm amide peak가 .

2-3 chitin chitosan

Chitosan chitin

chitin 가 amide 가 .

chitin chitosan 가 .

chitin chitosan

chitin chitosan Fig.12

, chitosan chitin

peak가 1530nm

2030nm peak가 . amide II

chitosan NH₂ peak . Chitin amide

peak가 chitosan

chitin chitosan

.

2-4 chitosan

Chitosan

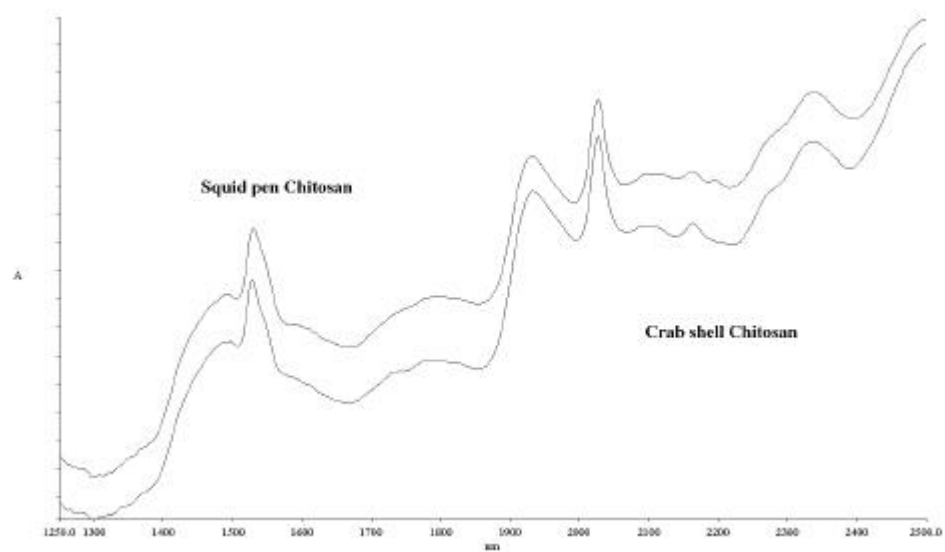


Fig. 11. Near-infrared spectra of crab shell chitosan and squid pen chitosan.

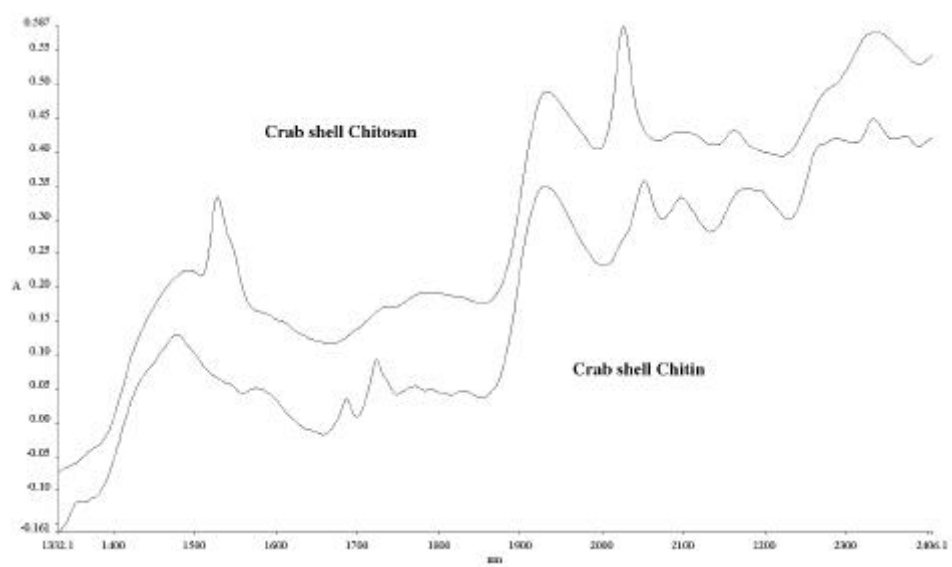
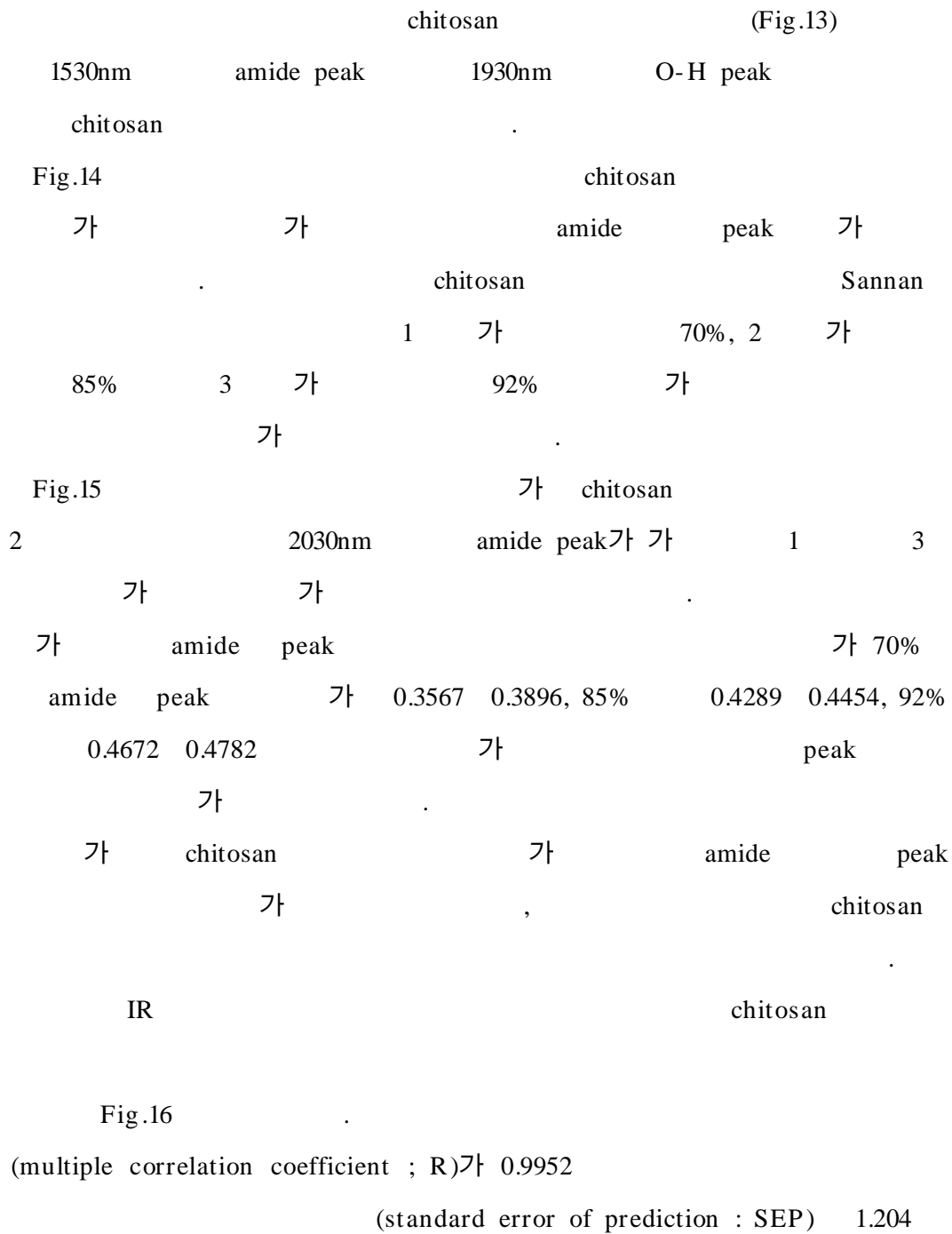


Fig. 12. Near-infrared spectra of crab shell chitin and chitosan.



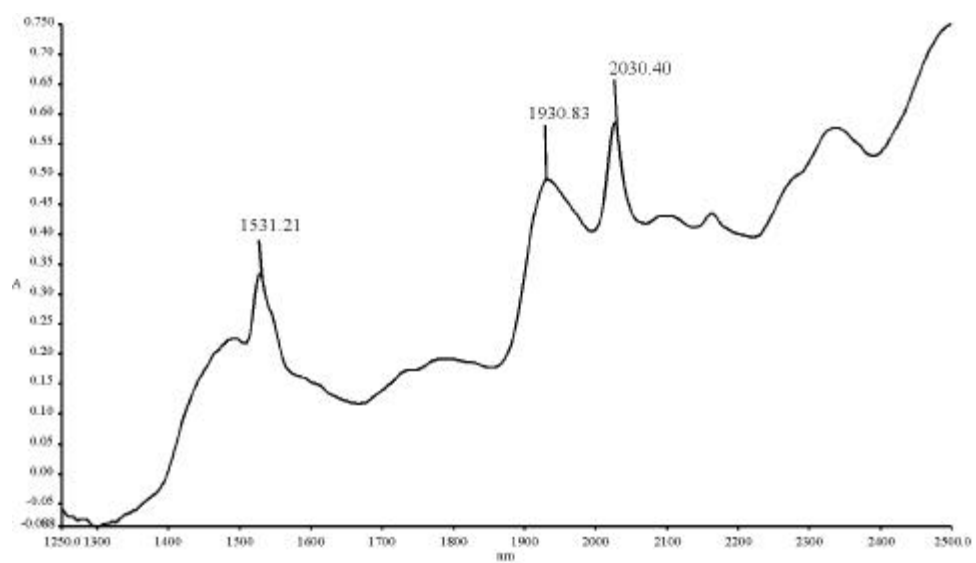


Fig. 13. Near-infrared spectra of typical chitosan.

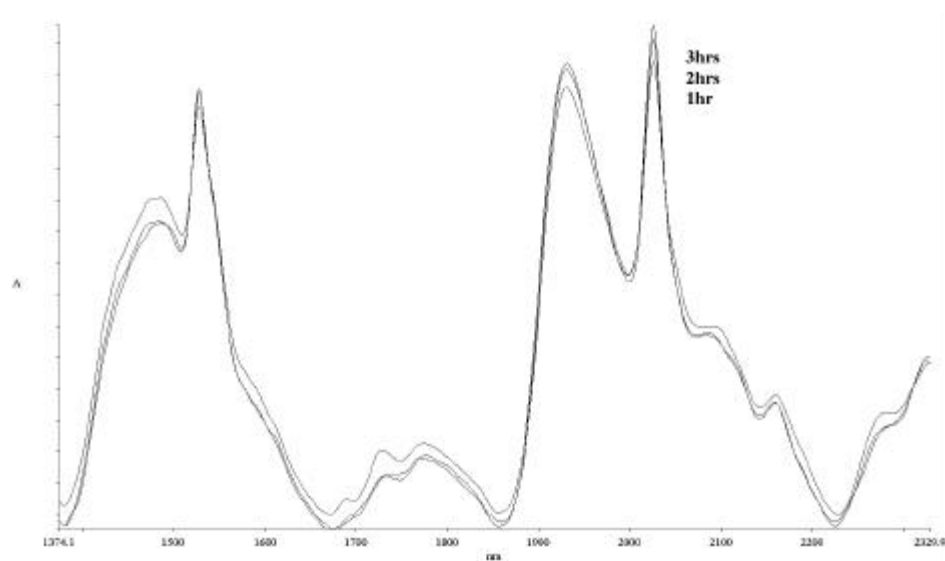


Fig. 14. Near-infrared spectra of different deacetylated chitosan.



Fig. 15. Second derivative near-infrared spectra of different deacetylated chitosan.

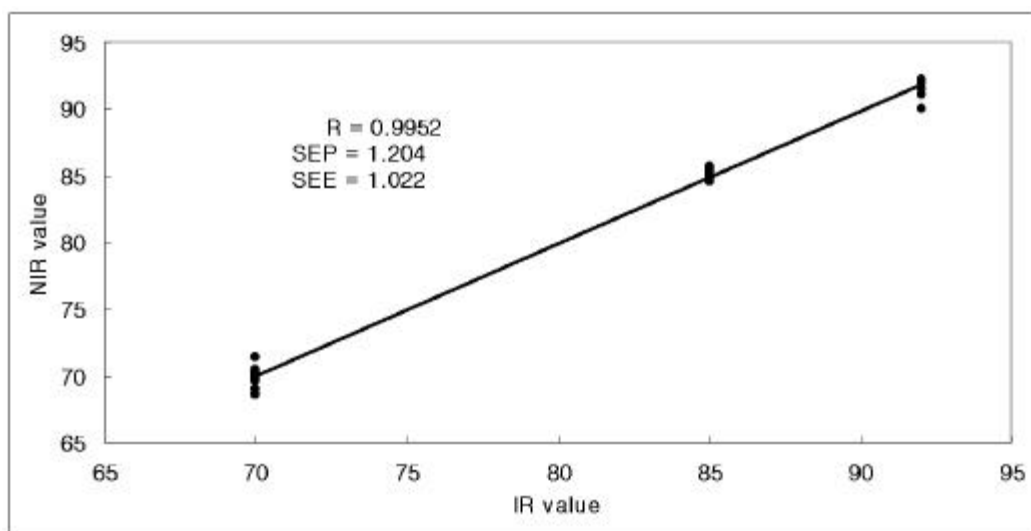


Fig. 16. Relationship between NIR method and IR method for determining the deacetylation degree of chitosan.

(standard error of estimate : SEE) 1.022 .

가 .

가 . (multivariate)

. Principal Component Regression(PCR)
 . PCR

scaling factor

. Principal Component(PC)

PC

PC가

가

, PC

. PCR

가

, PC

PC

가 chitosan

amide peak

R 0.9917

(SEP) 0.0069,

(SEE) 0.0061

(Fig.17).

chitosan

가

,

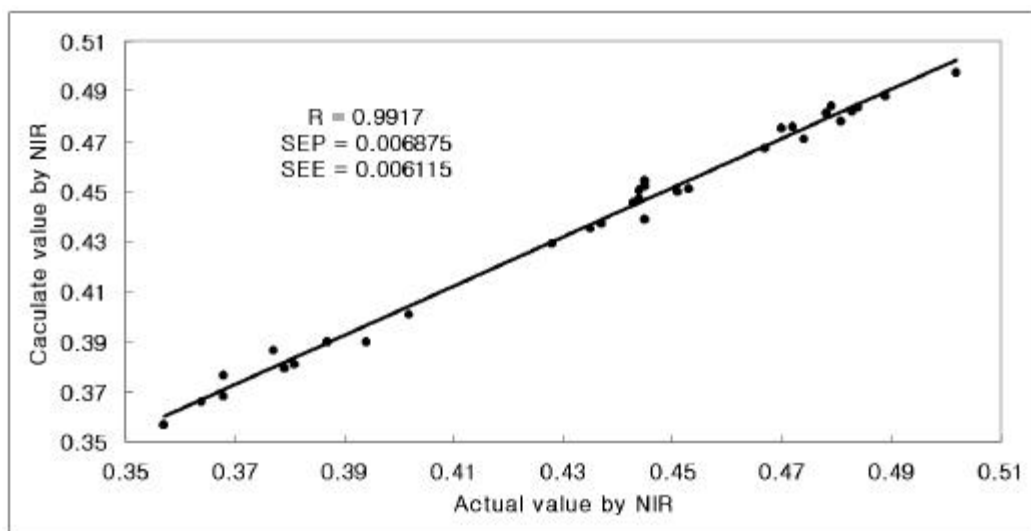


Fig. 17. Relationship between calculated deacetylation degree and actual deacetylation degree of chitosan by NIR.

chitosan

Table 4

2%

chitosan

2 - 5 chitosan

Chitosan chitosan amide (NH^{3+})

C-H

가

Chitosan

Fig.18 가 chitosan

chitosan chitosan

chitosan chitosan

peak가 1750 2150nm C-H

chitosan peak

chitosan amide

Fig.19 2 2030nm

amide peak가 peak가

Table 4. Deacetylation degree of chitosan by IR and NIR spectra

IR	NIR	Differences	NIR spectrum area
70	69.29	0.71	0.366
	69.96	0.04	0.3753
	70.17	-0.17	0.3794
	67.97	2.03	0.3567
	69.36	0.64	0.3607
	69.91	0.09	0.3896
	70.78	-0.78	0.3896
85	84.35	0.65	0.4289
	87.2	-2.2	0.4454
	85.04	-0.4	0.4369
	85.29	-0.29	0.4294
	86.09	-1.09	0.4387
92	91.68	0.32	0.476
	90.93	1.17	0.4754
	90.86	1.14	0.4782
	91.37	0.63	0.4709
	91.06	0.94	0.4672

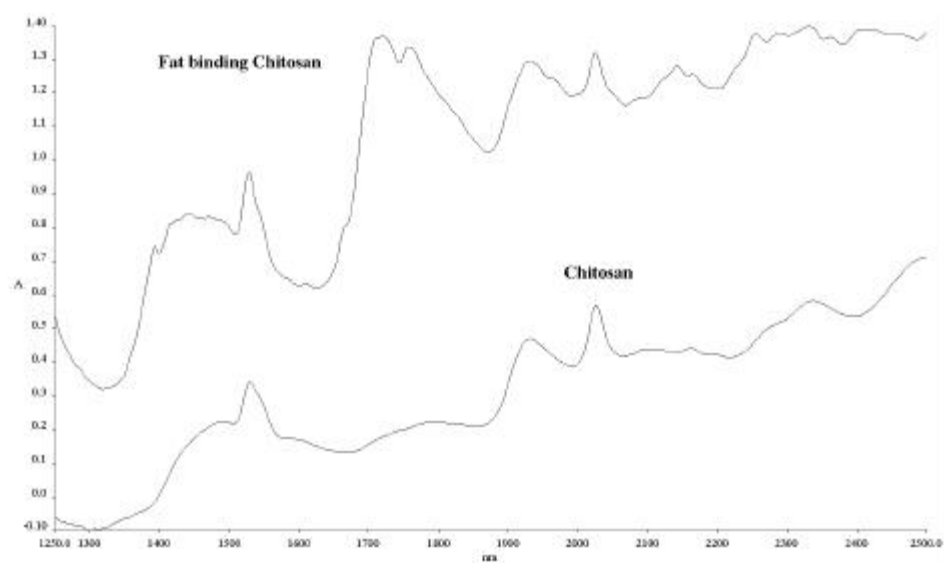


Fig. 18. Near-infrared spectra of chitosan and fat binding chitosan.

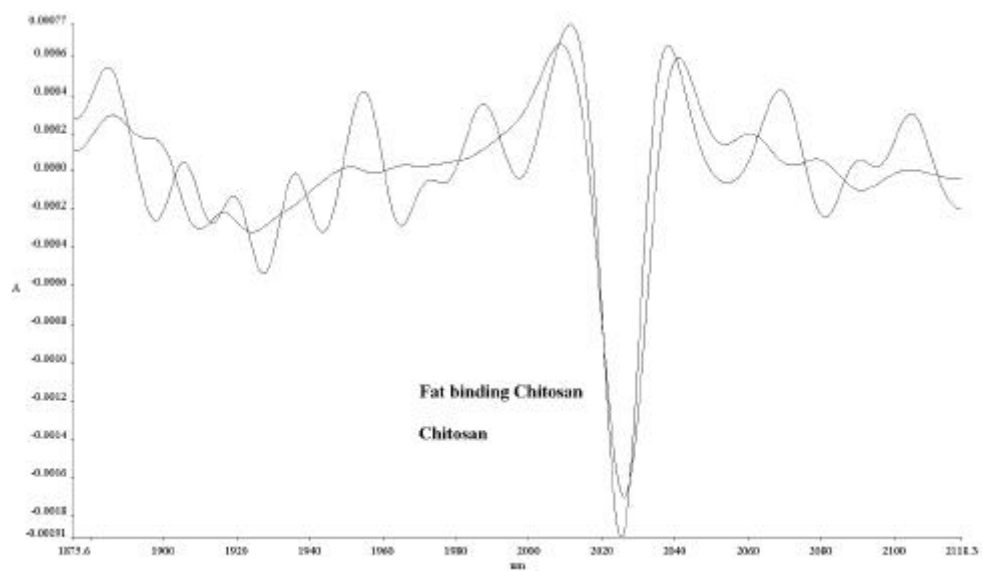


Fig. 19. Second derivative near-infrared spectra of chitosan and fat binding chitosan.

amide chitosan C-H
 chitosan 가

2-6 chitosan

Chitosan chitosan

Table 5 *in vitro* chitoasn

chitosan

Fig.20 Fig.21

in vitro chitosan 24kDa

chitosan 4kDa chitosan

24kDa

chitosan 4kDa chitosan

in vitro
 peak

Table 6 24kDa

chitosan 4kDa chitosan 659.3%,

338.8% chitosan 2 ,

peak 17.66, 12.82 chitosan

chitosan

Table 5. Degree of deacetylation and molecular weight of chitosan

		Degree of deacetylation (%)	Molecular weight
Chitosan^a	70H	70	28kDa
	70L		4kDa
Chitosan^b	85H	85	26kDa
	85L		4kDa
Chitosan^c	92H	92	24kDa
	92L		4kDa

^a Deacetylation ; added 47% NaOH to chitin for 1hr at 130

^b Deacetylation ; added 47% NaOH to chitin for 2hrs at 130

^c Deacetylation ; added 47% NaOH to chitin for 3hrs at 130

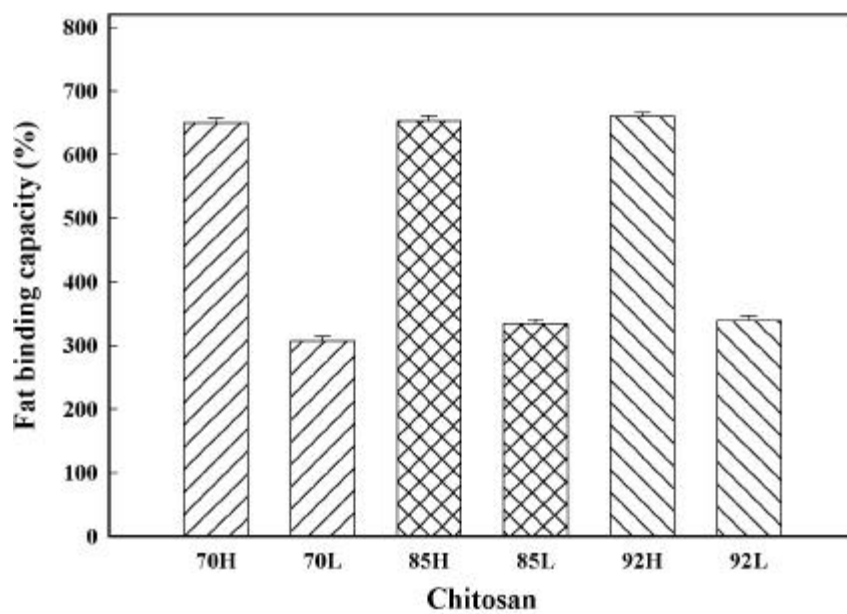


Fig. 20. Fat binding capacity of chitosan.

70H;DD70%, Mw28kDa chitosan, 70L;DD70%, Mw4kDa chitosan
 85H;DD85%, Mw26kDa chitosan, 85L;DD85%, Mw4kDa chitosan
 92H;DD92%, Mw24kDa chitosan, 92L;DD92%, Mw4kDa chitosan
 DD; Degree of deacetylation(%), Mw; Molecular weight.

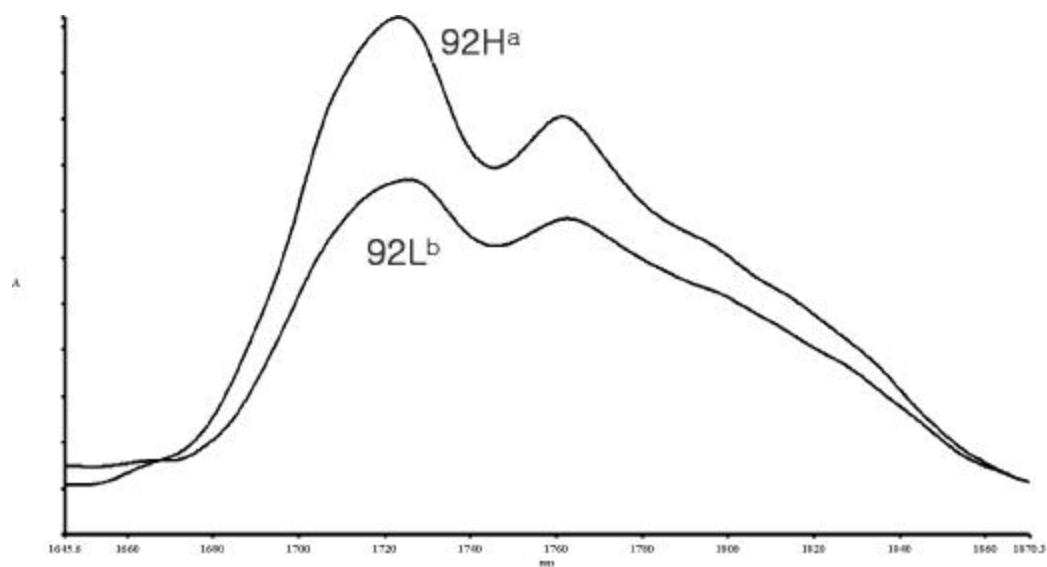


Fig. 21. Near-infrared spectra of fat binding chitosan.

^a92H; deacetylation degree(%) ; 92(%) Mw 24kDa

^b92L; deacetylation degree(%) ; 92(%) Mw 4kDa

Table 6. Fat binding capacity and corrected area by NIR analysis as molecular weight of chitosan

Chitosan	Fat binding capacity (%)	Corrected area by NIR analysis
92H^a	659.3 ± 6.8	17.66
92L^b	338.8 ± 8.5	12.82

^a92H; deacetylation degree(%) ; 92(%) Mw 24kDa
^b92L; deacetylation degree(%) ; 92(%) Mw 4kDa

가

chitin chitosan

가

chitin chitosan

chitin chitosan

chitin chitosan 가 (squid pen) (red snow crab shell) NIR 1200nm, 1510nm, 2050nm 2180nm peak가 peak

Chitin NIR

chitin

Chitosan chitin 가

amide 가 NIR

chitosan 1530nm, 2030nm NH₂ amide peak

chitin chitosan .

chitosan IR

NIR 가

가

가 1%

0.006

Chitosan NIR

C-H chitosan amide

, chitosan

Chitosan Mw 24kDa

chitosan Mw 4kDa chitosan

.

. ,
 .
 가 ,
 , , ,
 .
 .
 , ,
 .
 , , , ,
 .
Perkin Elmer , , ,
 .
 가 .
 .

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