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釜慶大學校 大學院

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金 孝 珍

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Application of UF membrane combined with pre-coagulation for DBPs precursor removal

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Abstract

Natural organic matter(NOM) which occurs ubiquitously in surface waters consists of humic (i.e., humic and fulvic acids) and nonhumic components. NOM in general as well as certain constituents is problematic in water treatment. The fractionation of NOM through water treatment processes can provide insight into treatment process selection and applicability. Recently, in water treatment industry, the problematic NOM fractions have been targeted for removal or transformation.

Membrane technologies have recently been extensively investigated as a water treatment process. When using the membrane filtration, the membrane, in general, can remove virtually all particles larger than the nominal pore size. Also, some membrane processes (e.g., Nanofiltration and Reverse osmosis) reject significant amounts of soluble species and are therefore prospective technologies for removing NOM.

However, these membranes are also easily fouled by NOM than other membranes (Ultrafiltration and Microfiltration). The concentration and type of NOM presented in a raw water will influence the characteristics of the water for

a particular membrane. Therefore, the purpose of pretreatment to a membrane process is to decrease the amount of irreversible fouling and increase the permeate flux. Applying coagulation process before membrane filtration has been suggested as means of reducing membrane fouling and improving the removal of dissolved organic materials that might otherwise not be removed by the membrane. Based on the results of this research, the following conclusions are summarized. For Nakdong river water, organic matter fraction represented 48.2% fraction of hydrophilic, 38.8% of fulvic acid, 13% of humic acid and 88% of dissolved organic matter has a molecular weight less than MWCO(Molecular Weight Cutoff) of 10K.

Therefore, removal of organic with only UF of MWCO 30K should not be expected. Also, applying coagulation process before membrane filtration showed not only increasing particle size but also improving the flux. That is, during the coagulation, substantial changes in particle size distribution occurred under rapid and slow mixing condition due to the simultaneous formation of microflocs and NOM precipitates. Flux decline on UF decreased with the increase in particle size due to the coagulation process (from raw water to slow mixing). Also, the flux decline is not influenced to pore size of membrane because particle more than regular size is formed by the simultaneous formation of microflocs and NOM precipitates from dissolved organic matters.

•

•

(THMs) ,

(Schnoor, J. L., 1979; Laine J.M., 1989).

1998 (phase 1) TTHMs

(MCLs: maximum contaminant levels) $80\mu\text{g}/\ell$, monochloroacetic acid, dichloroacetic acid, trichloroacetic acid, monobromoacetic acid dibromoacetic acid

HAA₅ $60\mu\text{g}/\ell$. 2002 (phase 2)

THMs HAA₅ $40\mu\text{g}/\ell$, $30\mu\text{g}/\ell$

• (membrane separation) DBP

(Taylor et al.,1990; Taylor et al., 1992; Jacangelo et al., 1989; Taylor, 1992; Amy et al., 1993; Siddiqui, 2000)

•

UF(Ultrafiltration), MF(Microfiltration), NF(Nanofiltration) 가 ,

•

MF/UF membrane 240 .

RO(Reverse osmosis)

•

• ,

• , 가 . RO

가

• 가 UF MF 가

(Conlon W. J. and Click J. D.,1984). UF

, virus, coliform

bacteria, *Giardia* cysts, *Cryptosporidium* oocysts

.

, fouling

.

UF

.

가 UF

. 1)

() 2)

fouling

() 3) ()

()

flux

(- -)

+UF

.

2.1

(pressure-driven)

(operated pressure), (pore size), (MWCO)

(Reverse Osmosis, RO), (Nanofiltration, NF), (Ultrafiltration, UF), (Microfiltration, MF)

Table 2.1. mechanism, pore size

. MF UF (sieve mechanism),

(ED) 가 , pore size

. table 2.1. pore size가

가 .

Table 2.1. Technically Relevant Main Membrane Operation

Membrane operation	Driving force(bar)	Mechanism of separation	Membrane structure (pore size)
Microfiltration	pressure (0.1- 2.0)	Sieve	Macropores(> 50nm)
Ultrafiltration	pressure (1.0- 5.0)	Sieve	Mesopores(2- 50)nm
Nanofiltration	pressure (5.0- 20)	Sieve+diffusion+size exclusion	Micropores< 2 μ m
Reverse osmosis	pressure (10- 100)	Diffusion +size exclusion	Dense (Macromolecular chains)
Electrodialysis	Electrical potential	Ion exchange	Ion exchange

1) (Reverse Osmosis, RO)

(RO) Fig. 2.1

, 가
10 100bar .

, 2
가 . 50 80bar 9
5 99% 가 가 (Mallevialle et al., 1996).

Reverse Osmosis

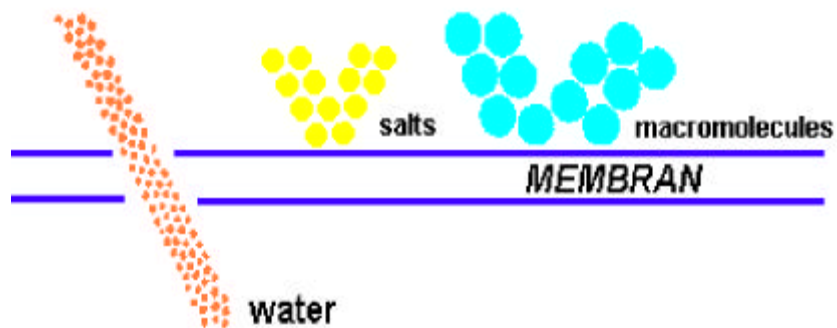


Fig. 2.1 Excluded species by Reverse osmosis.

2) (Nanofiltration, NF)

(NF)

(5 20bar)

, , , 가
(multivalent ion) (disinfection by-products)

(SOCs: synthetic organic compounds)

(Nanofiltration)

NF 가

(Sieving), (diffusion), (size exclusion)

, NF $10^{-3}\mu\text{m}$ 가

, RO $10^{-3}\mu\text{m}$ 가

1가 가

5 10bar (Low

Pressure Reverse Osmosis) 가

, 1가 20 70% 90%

200 500 10,000

(UF) (Cadotte et al., 1988).

3) (Ultrafiltration, UF) (Microfiltration, MF)

(UF)

. 10,000 100,000 가

가 Fig. 2.2

, 가 가 . MF , 가

. MF $0.1\mu\text{m}$

,

. UF 10^{-1} $10^{-2}\mu\text{m}$

가 , MF 10^{-1} $1.0\mu\text{m}$

.

, (UF) (MF)

MF가 0.1- 2.0bar , UF가 1.0- 5.0bar

. MF UF (sieving)

. MF UF ,

가 . 0.1NTU
(Olivieri et al., 1991; Laine et al., 1989;
Jacangelo et al., 1989, 1991).

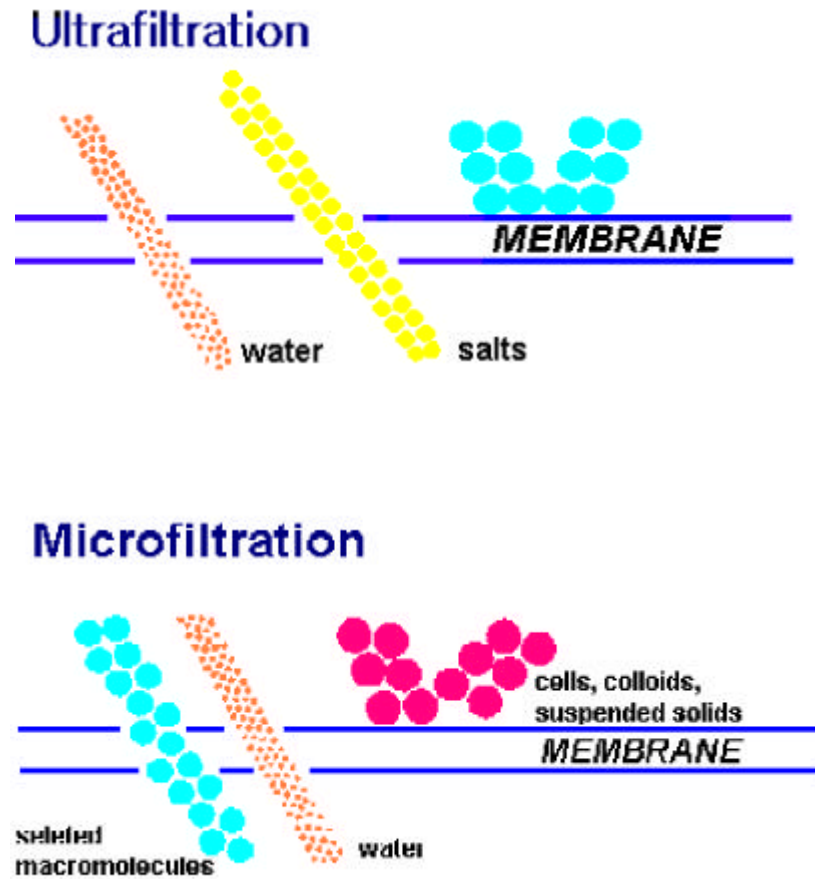


Fig. 2.2 Excluded species by Ultrafiltration and Microfiltration.

Fig. 2.3

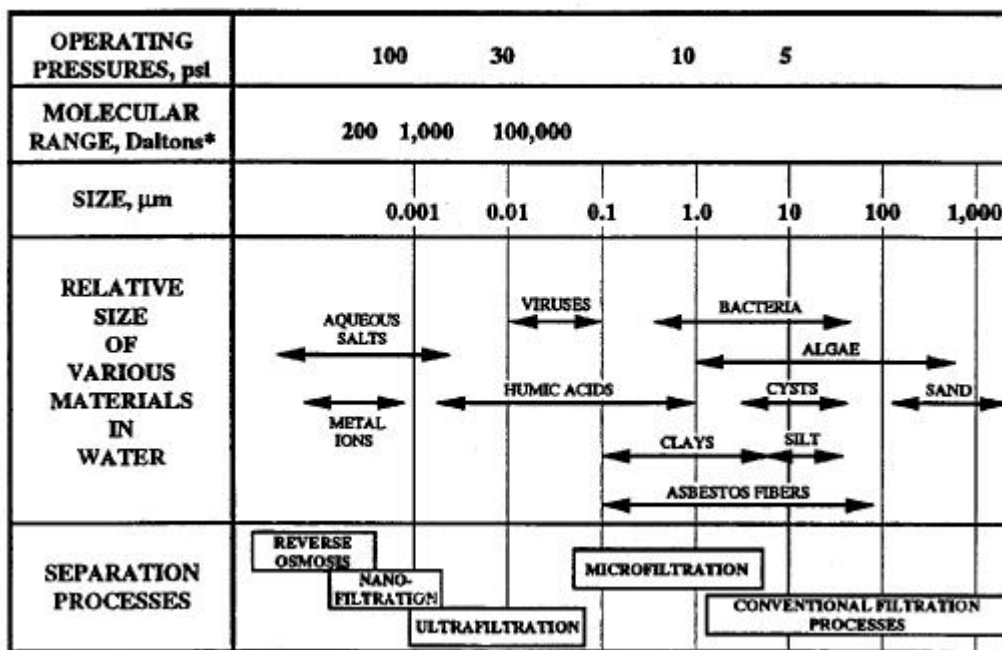


Fig. 2.3 Schematic comparison of selected separation processes.

Table 2.2. Reverse osmosis(RO), Nanofiltration(NF), Electro-dialysis reversal(EDR), Ultrafiltration(UF), Microfiltration(MF) 가

Table 2.2

(pathogen),	(organic solutes),	(inorganic solutes)
,	(TDS),	,
.	(DBPs)	
(SOCs)	.	mechanism size exclusion(sieving),
diffusion charge repulsion	가	. UF MF
MWCO가	,	.

Table 2.2. Characteristics of Membrane operation (ICR EPA, 1996)

process	Mechanism	Exclusion	Regulated solutes rejected by process		
			<u>Pathogens</u>	<u>Organics</u>	<u>Inorganics</u>
EDR	C	0.0001 μ m	None	None	Most
RO	S,D	0.0001 μ m	C,B,V	DBPPs, SOC	Most
NF	S,D	0.001 μ m	C,B,V	DBPPs, SOC	Some
UF	S	0.001 μ m	C,B,V	None	None
MF	S	0.01 μ m	C,B	None	Non

Mechanism: C=chage, S=size exclusion, D=diffusion

Pathogens: C=cysts, B=bacteria, V=viruses

Organics: DBPPs=disinfection by-product precursors, SOC=Synthetic Organic Compounds

2.1.1

가 가

.

(Plate and Frame),
(Tubular), (Hollow - fiber), (Spiral Wound)

,

Fig.

(1) (Tubular) (Hollow - fiber)
cylindrical geometry ,

Hollow fibers, 0.5 ~ 2.5mm
Narrow-bore tubes, 3 ~ 8mm
Wide-bore tubes, 10 ~ 25mm

Cabasso(1980) pore size

가 ,

10 .

가 ,

6 ~ 25mm , 13mm .

(surface area/volume ratio) ,

가

(2) (Spiral wound)

. 가
(feed spacer) 0.25 0.50 mm
, 1 NTU (Keily and Rogers, 1955).

(3) (Plate and Frame)

, Madsen(1977)
. UF 100 400m²/ m³ ,
5 20bar .

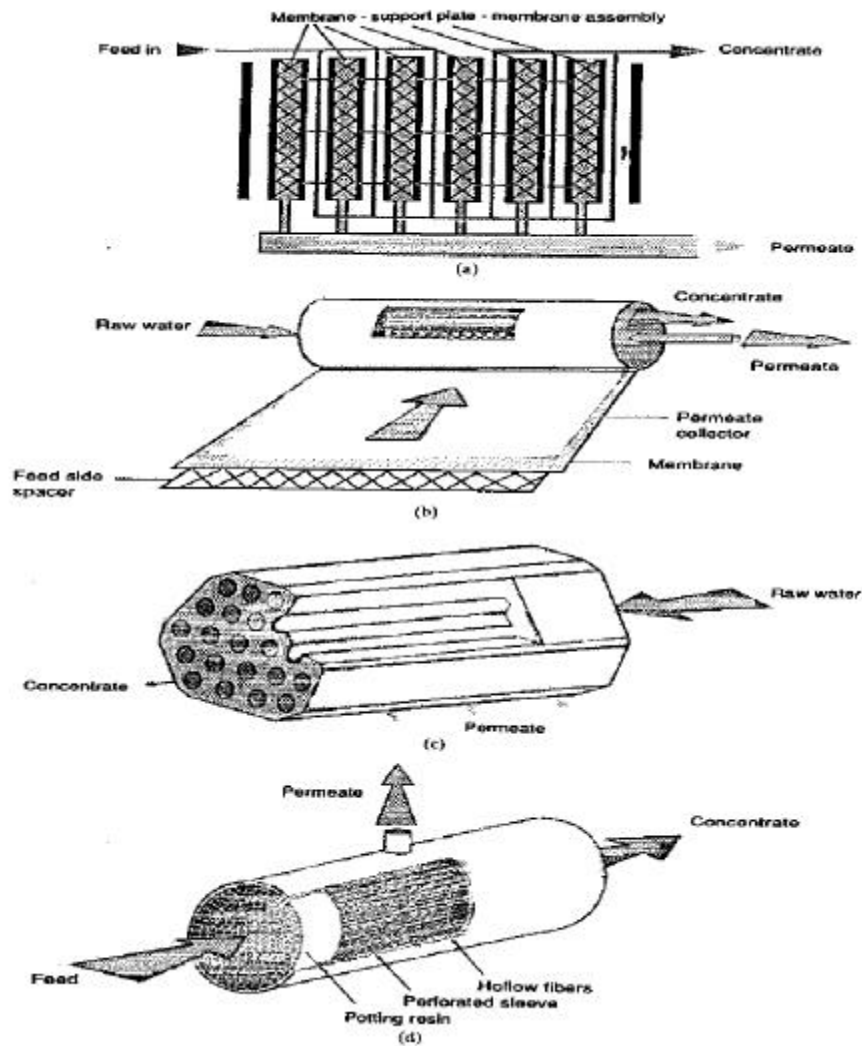


Fig. 2.4 Schematic representation of the four principal membrane modules: (a) plate and frame; (b) spiral module; (c) tubular module; (d) hollow fiber.

2.1.2

- (hydrophobic) (hydrophilic)
- . UF ,
- . UF MF ,
- 가 .
- ,
- (fouling) .
- .
- (1) Polysulphone(PS)
- 가 pH 1-13 가
- 1-20nm
- 가 ,
- 1,000-500,000 가 .
- (2) Polyamide(PA)
- (hydrophobic) ,
- .
- flux 가 .
- (3) Cellulose acetate(CA)
- 가 (hydrophilic) , (chlorine)
- , pH 4 7
- , ,
- (4) Ceramic

2MPa , flux
가 140 pH .

Fig. 2.5

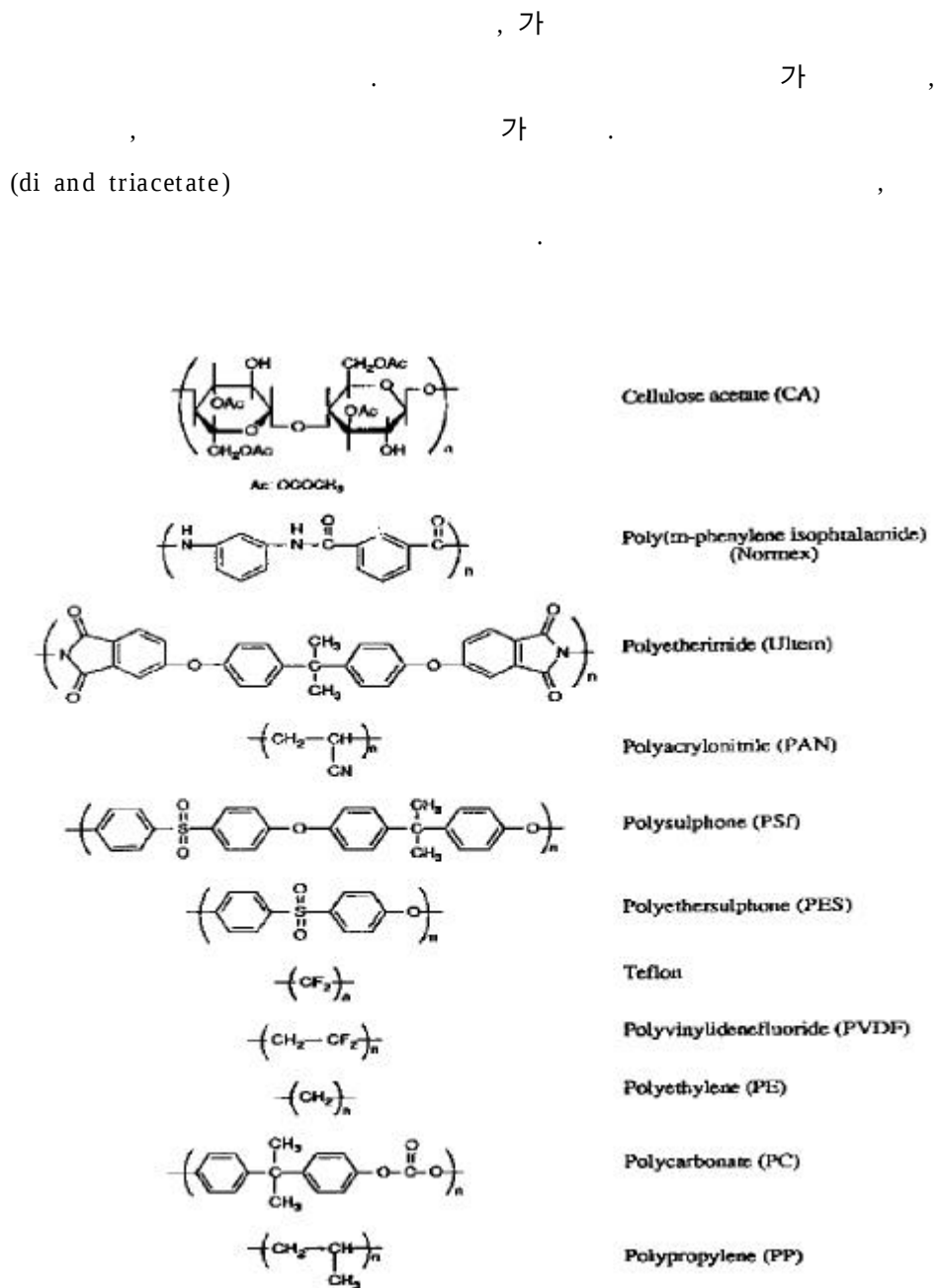


Fig. 2.5 Molecular structures of main organic membrane materials.

2.2

flux

MF UF

flux Darcy eq.

flux

$$J = \frac{\Delta p}{\mu R_m} \text{ ----- (2-1)}$$

Δp = (transmembrane pressure drop, TMP) [N/ m²]

μ = [N · s/ m²]

R_m = (hydraulic resistance) [1/m]

J가 J_s

.

$$J = \frac{(\Delta p - \sigma_k \Delta \Pi)}{\mu R_m} \text{ ----- (2-2)}$$

where, σ_k :

$\Delta \Pi$: [N/ m²]

, $\Delta \Pi$,

가 . , UF MF

,

.

UF MF

Poiseuille

(Laminar flow) . UF MF , r_{pore} ,

$$J = \frac{f r_{pore}^2 \Delta P}{8\mu\theta \delta_m} \text{----- (2-3)}$$

where, f : (open pore)

θ : (pore tortuosity factor)

δ_m : [m]

(2-3) 가 가 .

(MWCO)

가 , , f가 가 δ_m

(Resistance in series model) Sieve mechanism

가 . (J)

(ΔP) , (R_m) (R_f),

(R_c) (μ) (Cheryan,

1998). , δ_c cake(or gel) , k

(mass transfort coefficient) .

$$J = \frac{(\Delta p - \sigma_k \Delta \Pi)}{\mu(R_m(t) + R_c(\delta_c(t), \dots) + R_{cp}(k, J))} \text{----- (2-4)}$$

specific resistance . Kozeny equation
specific resistance .

2.3

Brownian diffusion

가 . MF UF

, .

Brownian diffusion ,

Shear-induced diffusion

. , D Brownian diffusion Shear-induced diffusion (Sethi and Wiesner, 1997).

$$D = \frac{kT}{6\pi\mu_0 r_p} + \frac{\tau_{wall}}{\mu_0 \eta(\phi)} r_p^2 D_{sh}(\phi) \text{ ----- (2-6)}$$

, τ_{wall} = (shear stress)

μ_0 =

$\eta(\phi)$ =

Brownian Shear-induced diffusivity μm 가 particle

back transport가 가 flux가 가 .

Fig. 2.6 Brownian

diffusion Shear-induced diffusion , D가 $10^{-1} \mu m$ size

가 . ,

가 가 가 .

가

(Fane, 1984; Wiesner, Clark, and Mallevialle, 1989; Lahoussine-Turcaud et al., 1990).

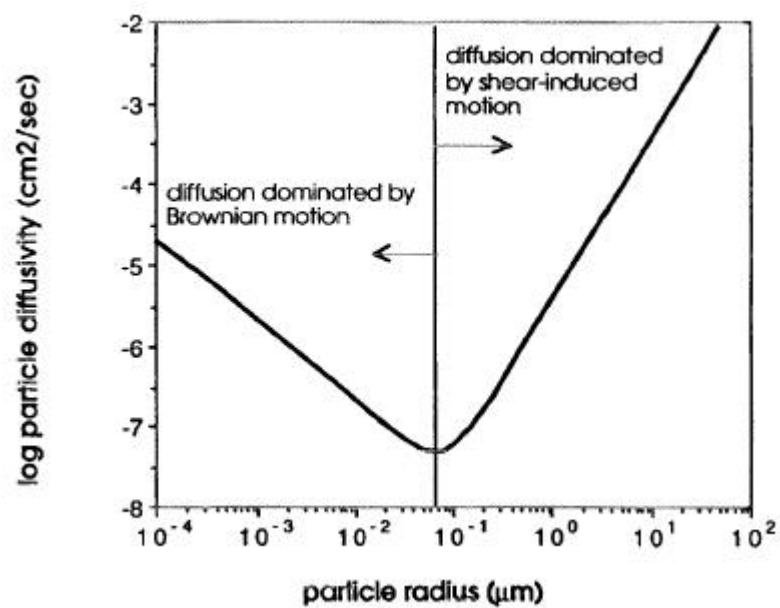


Fig. 2.6 Brownian and shear-induced diffusivity as a function of particle size for condition typical of hollow fiber UF membranes.

2.4 (Seperation Mechanism)

가
(rejection) . UF MF (size exclusion) 가
, RO

Global rejection 1 (permeate water conc., C_p) (feed
water conc., C_f) .

$$R = 1 - \left(\frac{C_p}{C_f} \right) \text{-----} (2-7)$$

Local rejection

$$R_{local}(x) = 1 - \left(\frac{C_p(x)}{C_{mem}(x)} \right) \text{-----} (2-8)$$

, C_{mem} : [mg/ ℓ]

$C_m = C_{mem}(x) = (PF) C_{bulk}(x)$, C_m (module)

가 , bulk
가 .

$C_{wall} = (PF) C_{bulk}$.

R_{local} .

C_{wall} Bulk C_{bulk} $R_{apparent}$ 가 .

$$R_{apprent} = 1 - \frac{C_p}{C_{bulk}} = 1 - (1 - R_{local})(PF) \text{ ----- (2-9)}$$

	R	module	local rejection
가	가		rejection mechanism
	.		

2.4.1

UF MF , , (dispersion forces), (hydrophobic bonding) 가 , humic, fulvic acids . Mechanical sieving(Steric Rejection) 1936 Ferry 가 Fig. 2.7 R P , (lag velocity) . (1- P) , $\lambda = r_p / r_{pore}$ (2-9) .

$$p = \begin{cases} (1 - \lambda)^2 [2 - (1 - \lambda)^2] G & \lambda \leq 1 \\ 1 & \lambda > 1 \end{cases} \text{----- (2-10)}$$

G Zeman Wales(1981) (Lag coefficient) .

$$G = \exp(-0.7146 \lambda^2) \text{----- (2-11)}$$

Lakshminaryaniaah(1965) G

$$G = 1 - 2.104\lambda + 2.09\lambda^2 - 0.95\lambda^5 \text{ ----- (2-12)}$$

$$(2-10) \quad (2-11)$$

가 .

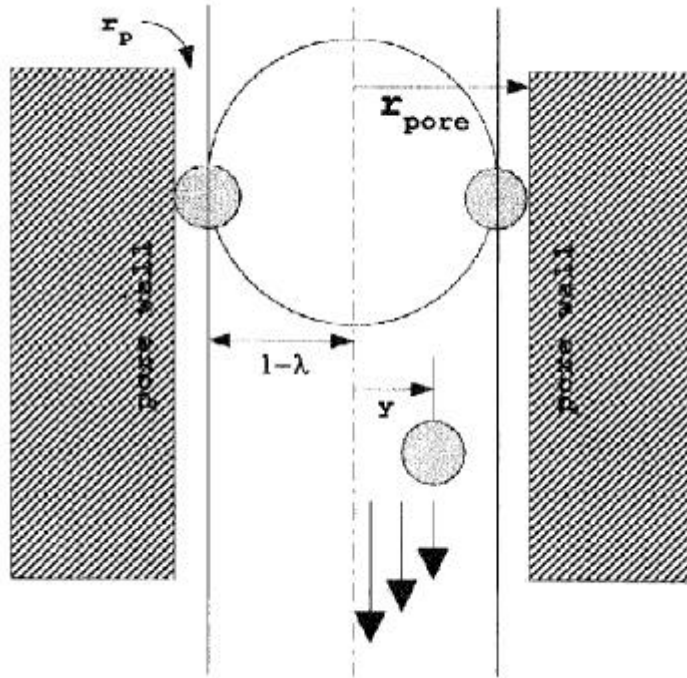


Fig. 2.7 Definition of particle removal in a membrane pore.

$$R_{apparent} = \frac{(1 - p)}{PF} \quad R_{local}^l \quad p^*$$

(Deen, 1987).
(Davidson and Deen, 1988).

$$a_p = Z_1 (\overline{M})^{Z_2} \text{-----} (2-13)$$

$$Z_1, Z_2, \dots, Z_2 \quad \text{가}$$
$$Z_2 \quad 1/3$$

가

NOM 가

UF	NF
1	1
2	2
3	3
4	4
5	5
6	6
7	7
8	8
9	9
10	10
11	11
12	12
13	13
14	14
15	15
16	16
17	17
18	18
19	19
20	20
21	21
22	22
23	23
24	24
25	25
26	26
27	27
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32	32
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77	77
78	78
79	79
80	80
81	81
82	82
83	83
84	84
85	85
86	86
87	87
88	88
89	89
90	90
91	91
92	92
93	93
94	94
95	95
96	96
97	97
98	98
99	99
100	100

(Taylor et al., 1988; Thompson, and Carswell, 1987). NOM

THMs

. THM

. DOC

NF

GAC

DOC

2.4.2

가 가
가 가 가
가 (functional group)가 NF
RO 가
가
SOCs
THMs,
VOC RO 가
가

2.5 (Concentration polarization)

(concentration polarization) (fouling)
가
가
(convective flow)
가
가
가
1) (osmotic pressure) 가 flux
2) 가 flux
3) 가
(pore plugging) flux가

- 4) .
- 5) , ,
water flux fouling (Clark, 1994).
가
. Fig. 2.8
.
(fouling)
- flux mechanism .
- 1) (gel polarization).
2) cake (concentration polarization layer) 가
.
3) / .
4) flux가
가 , /
.
가 (hydrophobic) pH, ,
(isoelectric point)
(Catherine J., Mark M. Clark, 1994).

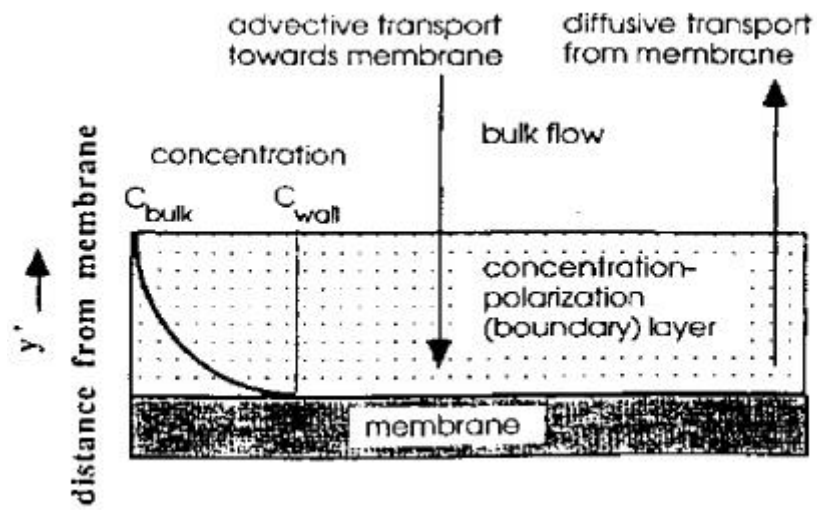


Fig. 2.8 Changes in solute concentration within the concentration polarization layer.

Fig. 2.9

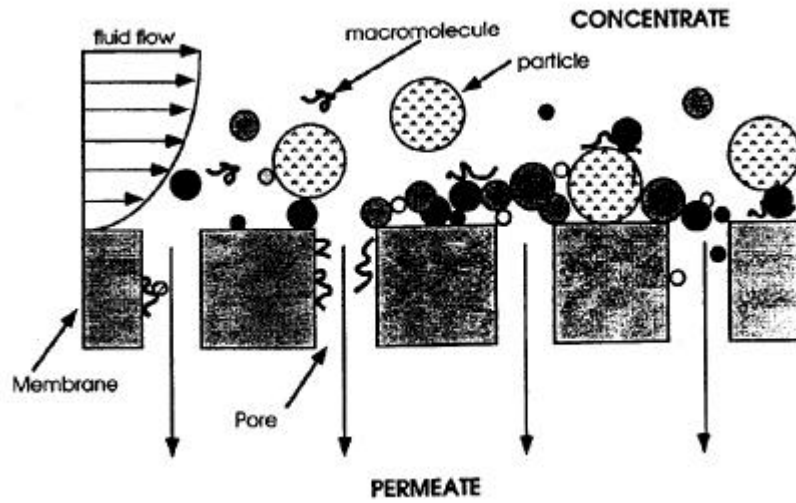


Fig. 2.9 The accumulation of materials on, in, and near a membrane in the presence of a cross flow.

2.6 (NOM)

2.6.1

Humic substance 가

(MacCarthy and Suffet, 1989). Humic Substances

lignins tannins (condensation) phenolic polymers가

.

. Humic substance 가

,

.

humic substance pH fulvic acid,

humic acid humin . Fulvic acid pH ,

humic acid (<pH 2) . Humin pH

. Fulvic acid 500 2,000 dalton

(Thurman et al., 1982). humic acid 2,000 dalton 가

. Humic substance .

humic substance 가

.

soil humus peat

.(Hall and Packham, 1965)

(DOC)

allochthonous . humic substance cellulose lignin

(Malcolm, 1985). Allochthonous

humic substance가 , , autochthonous

humic substance (Steinberg and Muenster,

1985). Autochthonous humic substance

(Malcolm, 1985). Hemicellulose protein , protein chitinous

substance humic substance .
 Humic substance .
 humic substance ,
 (MacCarthy and Suffet, 1989). Humic substance
 , aluminum iron (Thurman et al.,
 1982).

2.6.2

Humic substance pH
 . humic substance humic substance
 , Table 2.1 humic substance
 .

Table 2.3 Humic substances classification based on solubility

Current designation	Solubility Characteristics
Humic acid	Soluble in alkali, precipitated by acid
Brown humic acid	Not coagulated from alkali solution in the presence of electrolyte
Gray humic acid	Coagulated in the presence of electrolyte
Fulvic acid	Soluble in alkali, not precipitated by acid
Hymatomelanic acid	Soluble in alkali, precipitated by acid, soluble in alcohol
Humin	Insoluble in alkali

humic substance fulvic acid(pH)
 humic acid(pH 2), humin(pH
) . Fulvic acid ,
 (Leenheer, Brown, and Noyes, 1989).

fulvic acid (Wilson, 1959), fulvic acid humic substance 가
 , humic acid
 . Fulvic acid humic substance 90%
 (Malcolm, 1985), 10% humic acid .
 NOM humic substance(humic acid, fulvic acid)
 nonhumic substance(, , 가)
 .(Douglas et al., 1993; Edward et al., 1995).

Table 2.2 . , NOM humic substance nonhumic substance
 , humic substance
 가 nonhumic substance(,)
 .

Table 2.4 Characterization of bulk NOM

Fraction		Species
Humic NOM (Hydrophobic)	Humic acid Fulvic acid	Hydrophobic acids
Nonhumic NOM (Hydrophilic)	Hydrophilic acids Biochemical	Amin acids Proteins Carbohydrates

NOM ,
 , , ,
 ,
 , 가 . (Bruchet and Ryback, 1996)
 Table 2.3 NOM
 .

Table 2.5 Natural organic matter fractions and chemical groups
(Thurman and Malcolm, 1983)

Fraction		Chemical Groups
Hydrophobic	Acids	humic · fulvic acids, aromatic acids, phenols ...
	Bases	proteins, aromatic amines ...
	Neutrals	hydrocarbons, aldehydes, ethers ...
Hydrophilic	Acids	sugars, sulfonics, hydroxyl acids ...
	Bases	amino acids, purines, pyrimidines ...
	Neutrals	polysaccharides, aldehydes, ketones ...

Humic substance, carbonyl, phenolic, alcoholic, hydroxyl, carboxyl, methoxyl functional group 가 (Edward et al., 1995). Humic acid fulvic acid 가 polymetic compounds . 가 , pH protonation deprotonation .



deprotonation 가 (polyanions) . Humic fulvic acid . , (10- 15 μeq/mgC at pH 8.0) 가 fulvic acid (5- 10 μeq/mgC at pH 8.0) 가 humic acid

. hydrophilic acids pH 8 45 μ eq/mgC
 가 가 .
 가 가 .
 humic substance ,
 DOC humic nonhumic substance , humic substance
 , 가 . Nonhumic substance
 (ex, hydrophilic acids, proteins, amino acids, carbohydrates) ,
 (Douglas et
 al., 1993). , nonhumic humic
 DBP_s . , nonhumic
 BDOC .
 . , humic
 nonhumic 가 (Leenheer, 1985).
 nonhumic(hydrophilic)
 45% .
 NOM
 .
 , lignin
 .
 lignin . lignin
 , NOM NOM
 (Edward et al., 1995).

2.6.3 Humic substance

가 가 .

. Total Organic Carbon(TOC), Dissolved Organic Carbon (DOC), UV absorbance(UV_{abs}), Assimilable Organic Carbon(AOC), Biodegradable Dissolved Organic Carbon(BDOC) ,

Apparent Molecular Weight Distribution(AMWD)

Gel Permeation Chromatography(GPC), Ultrafiltration, X-ray .

(DBP_s) .

, THM Trihalomethane Formation Potential(THMFP), Haloacetic acids(HAAs), Total oxiganic halogen(TOX) 가 (Miltner, 1996).

Humic acid fulvic acid, hydrophilic acid (XAD-8 XAD-4)

. , Humic substance XAD-8 , hydrophilic acid

XAD-4 가 . hydrophilic acid

fulvic acid sugar amino sugar가 fulvic acid

humic acid 가 가

가 . Humic acid fulvic acid

가 . Humic acid C, H, O, N, S, P ,

50% , fulvic acid . O, N, S

(,) ligand .

Figure 2.1 2.2 fulvic acid humic acid .

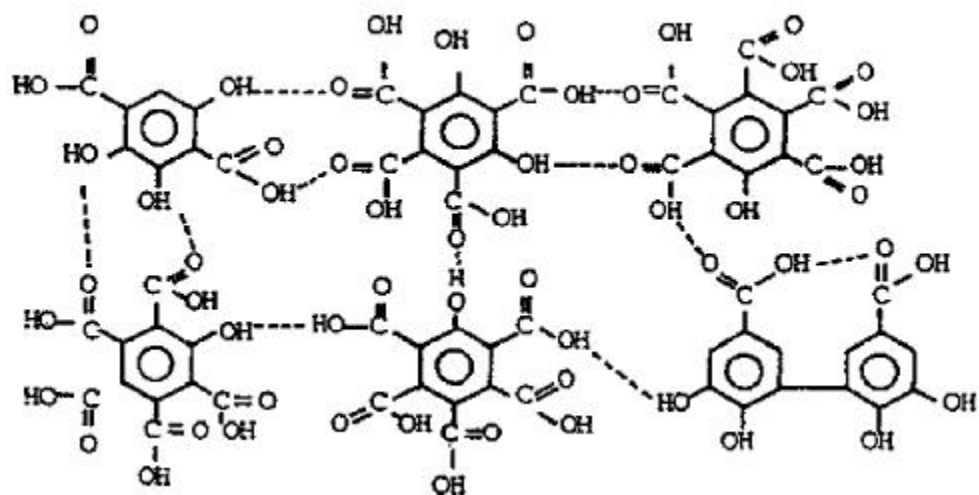


Fig. 2.10 Structure of humic acid.

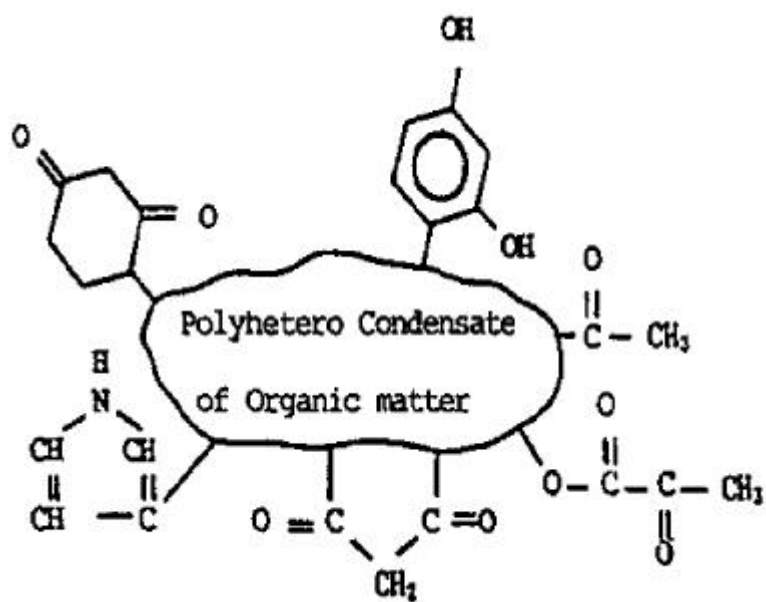


Fig. 2.11 Structure of fulvic acid.

2.7

(Chlorination)

가

가

THMs(trihalomethans)

(Rook 1974)

THM

Fig. 2.12

가

NOM (Natural Organic Matters)

가

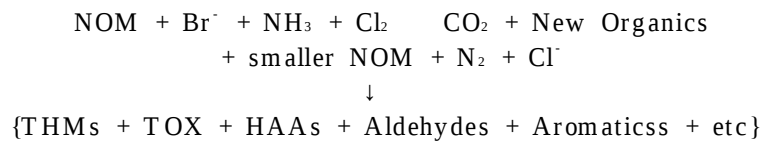
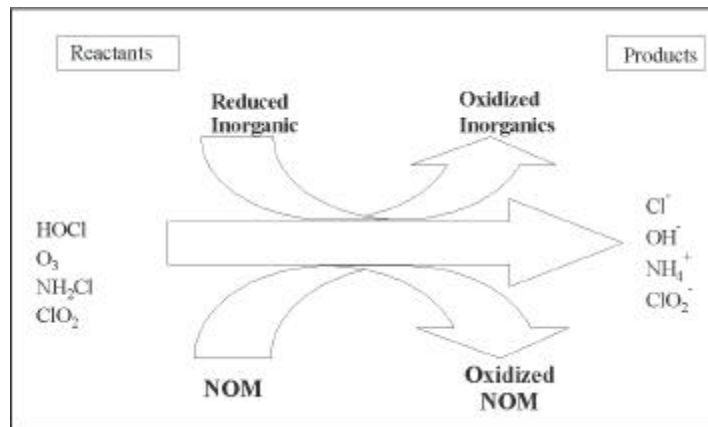


Fig. 2.12 Schematic illustration of reaction of various oxidants with natural organic material and reduced inorganic substances.

(USEPA)

(NAS, 1977)

1998 (phase 1)

DBPR TTHMs

(MCLs maximum contaminant levels)가 $80\mu\text{g}/\ell$, monochloroacetic acid, dichloroacetic acid, trichloroacetic acid, monobromoacetic acid dibromoacetic acid
 HAA_5 $60\mu\text{g}/\ell$. bromide ion

$10\mu\text{g}/\ell$, chlorite ion $1\text{mg}/\ell$. 2002 (phase 2)
 DBPR(disinfection by-products rule) THMs HAAs가 $40\mu\text{g}/\ell$, $30\mu\text{g}/\ell$. Rook(1974) Rotterdam waterworks
 headspace THM . chloroform, dichlorobromomethane, dibromochloromethane, bromoform 4가
 (haloforms) ,
 . Rook(1974) THMs
 THMs .
 Rook(1974) 가 가
 bromohaloform ,
 HOCl(hypochlorous acid) bromide HOBr(hypobromous acid) (aromatic)
 .
 Table 2.6 Croue Martin(1991) DOC
 THMFP TOXFP(Total organic halide formation potential)
 Humic acid DOC
 THMFP TOXFP 가
 . Hydrophilic acids DOC THMFP
 , DOC TOXFP TOXFP
 hydrophilic .

Table 2.6 Present the THMFP and TOXFP of the different extracts
(J. P. Croue, B. Martin, 1991)

Fraction	UV ₂₅₄ :DOC (m ⁻¹ :mg)	Chlorine Demand (mg Cl ₂ /mg C)	THMFP (μg CHCl ₃ /mg C)	TOXFP (μg Cl/mg C)
Humic acids	4.6	3 ± 0.2	51 ± 2	277 ± 34
Fulvic acids	3.1	1.4 ± 0.12	26 ± 2	140 ± 5
Hydrophilic acids	2.0	1.2 ± 0.2	21 ± 1.4	101 ± 12
Hydrophobic neutral	2.0	0.27	12	40

Fig. 2.13

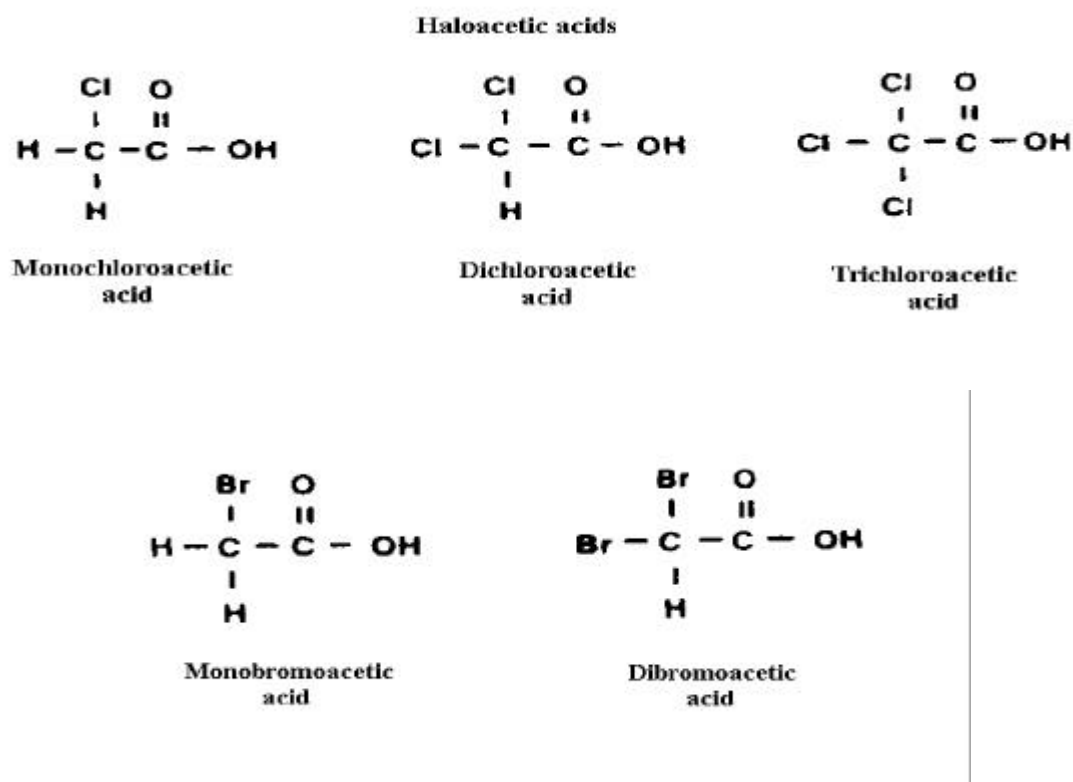
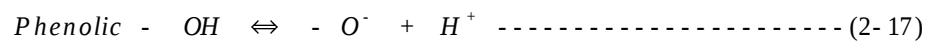
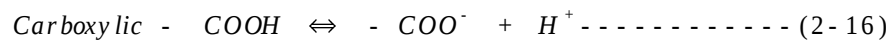


Fig. 2.13 Structural formulas for DBPs(Krasner S. W., 1989).

2.8

pH ,
 가 pH
 가 (2- 16, 2- 17). pH
 가 가
 가 가
 fulvic acid 가 humic acid
 . Narkis Rebhun(1977)



mechanism
 . 가
 mechanism
 mechanism . 3가
 mechanism , sweep (Gregor et
 al., 1997). 가

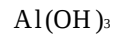
sweep coagulation
 가 . Al(OH)_3
 가 .
 floc sweep .
 Al() Fe()
 가 . 가
 가 .
 mechanism , Fig. 2.13

가 monomeric polymeric aluminum complexes
 HA-Aluminum
 Aluminum hydroxide

Fig. 2.14 Al() 가

$Al(OH)_3(s)$
 monomeric Al() polymeric Al() $Al(OH)_3(s)$
 Al()-
 $Al(OH)_3(s)$
 pH (Hall and Packham, 1965; Edzwald et al., 1977). Hall Packham (1965) Edzwald (1977) Alum
 pH 5 6.5
 Mangravite (1975) 5mg humic acid/L
 pH 가 가
 가 가Al
 () 가 humic acid

mechanism
 pH
 4 5.5 mechanism pH 가
 가
 mechanism sweep coagulation
 Al(OH)₃가 pH 6 8 Al(OH)₃
 mechanism
 가 sweep coagulation mechanism



가

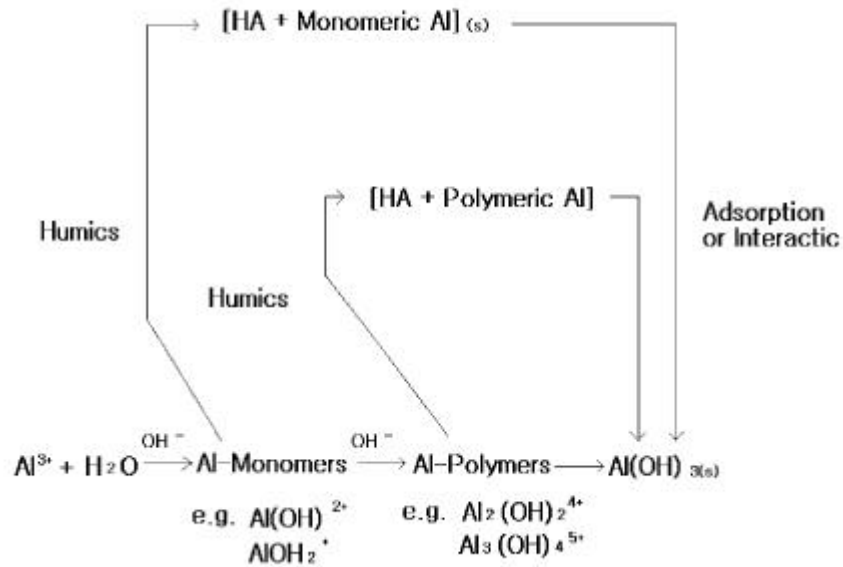


Fig. 2.14 Mechanism for alum coagulation of humic substances(Dempsey, 1990).

3.1

UF Fig. 3.1

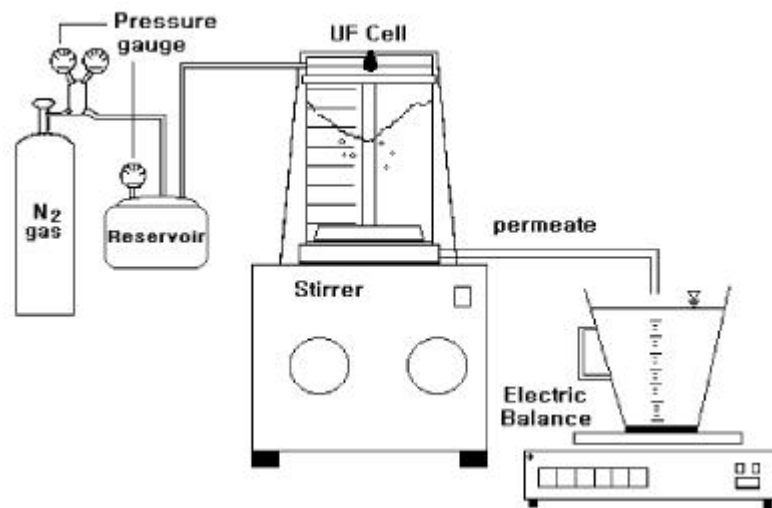


Fig. 3.1 Experimental schemetic of ultrafiltration membrane.

UF Millipore batch
 400M $\emptyset$, (Diameter) 76mm , Dead-end UF Membrane stirred cell .
 hydrophilic , (Molecular weight cutoff: MWCO)
 30,000Daltons YM30 .
 magnetic stirrer
 가
 , flux (L/ hr $\cdot$ m²) .
 75psi (5.3kg/cm²)
 42.69psi(3kg/cm²) .
 glycerin

30 1 skin(glossy)
 1 (backflushing) disc
 (skin) 14.23psi(1kg/ cm²) 10%
 (25) .

3.2

3.2.1 (Organic fraction)

DOC가 (1,000 mg/L)
 (Thurman et al., 1981). cleaning
 process Standard Method(19th Edition, 1995)

0.1N NaOH 24 가 .
 Soxhlet hexan, methanol, acetonitrile 24
 Soxhlet .

0.1N NaOH .
 , 0.1N NaOH, 0.1N HCl DOC가 0.5 mg/L
 , (Thurman et al., 1981;
 Daignault et al., 1988; Leenheer, 1981).

bed 1 bed volume . ,
 4mL/ min .

$$P = \frac{C_p}{C_r} \text{----- (3-1)}$$

where, C_p :

C_r :

P - 가 . Cutoff-size
, 가 (,
)
가 .
 C_{ro} MWC AMWD .
MWC C_{ro} AMWD . Logan-
Jiang P C_{ro} .

$$\ln C_p = \ln (P \cdot C_{ro}) + (P-1) \cdot \ln F \text{----- (3-2)}$$

where, C_p :

F : 1- (/)

P C_{ro} $\ln(C_p)$ $\ln(F)$ plot . C_{ro}
.

$$C_{<j,i>} = \frac{C_{ro(iMW)} - C_{ro(jMW)}}{\text{Initial sample concentration}} \text{----- (3-3)}$$

, 0 가 P MWC 가 DOC 가
. Logan Jiang $P < 0.2, P > 0.9$ AMWD
가 . P C_{ro}
 C_{ro} .

3.2.3 THMFP (Trihalomethane Formation Potential)

THM THMFP GC
liquid-liquid method(-),
Head-space, purge & trap .
가 , ,
Head-space , .
Standard Method , THMFP
, 가 U.S.
EPA (UFC : Uniform Formation Conditions., Scott R., 1996)
,
.

pH 8 borate buffer 2.0 mL/L 가 .
, H₂SO₄/NaOH pH 8 ± 0.2 .
incubation bottle 3/4가 .
hypochlorite-buffer solution 24hr 가 1.0 ± 0.4 mg/
L .
가 , , bottle cap
2 .
headspace가 가 .
10 .
24h 20.0 incubator .

standard method (Na₂S₂O₃ · 5H₂O)
0.1mL가 5mg
Gas Chromatography .

Table 3.1 Analytical condition of GC/ECD

Item	Condition
Injector Temp.	200
Detector Temp.	220
Initial Temp.	60
Initial Time	15min
Final Temp.	60
Total Flow	12.24 mL/min
Column Flow	0.61 mL/min
Gas	N ₂
Detector	ECD
Column	5MS (Crosslinked 5% PHME Siloxane, 30m × 0.25mm × 0.25μm)

3.2.4 TOXFP (Total Organic Halide Formation Potential)

0.5% Na₂SO₃

H₃PO₄ (1+10) 4 5 가 pH 2 25mL TOXs

(AD-2000, Dohrmann) 400mg (ψ 7mm × 5

cm) 1mL/min Total halide KNO₃ 8.2g 1L

Nitrate solution 5mL (1mL/min) inorganic halide

Organic Halide Analyser (DX-2000,

Dohrmann) TOXs . Carrier gas

, 200mL/min halide (80

0 850) 가 gas titration cell Ag

3.2.5

. 6 Phipps and Birds Jar - tester
 , paddle $(7.5\text{cm})^W \times (2.5\text{cm})^L$.
 tachometer가 .
 Jar $(11.5\text{cm})^W \times (11.5\text{cm})^L \times (21\text{cm})^H$ 2L Jar
 . 10cm
 . $\text{Al}(\text{OH})_3$ $\text{Al}_2(\text{SO}_4)_3 \cdot 16\text{H}_2\text{O}$
 $\text{Fe}(\text{OH})_3$ FeCl_3 .
 0.25M(as alum) stock solution
 10g/L(as alum) dosing solution . Dosing
 solution 가 dosing solution
 .
 (rpm) (G)
 . G
 250rpm ($G=550\text{sec}^{-1}$ at 25 , $Gt = 33,000$) 30rpm ($G=22 \text{ sec}^{-1}$ at 25 ,
 $Gt = 39,600$) UF . 5min, 30min
 30min
 , 10cm .
 .

3.3

(1) UV₂₅₄ (UV 254nm absorbance, (1/cm or 1/m))

(Ezdwald et al., 1985). lignin, tannin, humic aromatic substance aliphatic substance 200-400nm 가 . pH 4 10 가 pH 7 pH NaOH HCl cell Type A/E Glass Fiber Filter(Gelman Science) 1μm cell 254nm .

(2) (turbidity)

HACH, 2100P turbidimeter , 0-4000 NTU 1 (formazin) , 2 . 가 .

(3) TOC(Total Organic Carbon)

TOC , TOC vial . TC standard C₈H₅KO₄(anhydrous potassium biphthalate) IC standard Na₂CO₃(anhydrous sodium bicarbonate), NaHCO₃(anhydrous sodium bicarbonate) TC IC .

(4) DOC(dissolved organic carbone)0.45 μ m

membrane filter TOC

(5) (alkalinity)

가 가

0.02N

pH 4.5

ml

CaCO₃

Table 3.2 Analytical method and instruments

Item	Unit	Analytical method and instruments
Jar-test	-	Jar tester (Phipps & Bird, Model 7790-500)
pH	-	pH meter (METTLER DELTA 345)
Turbidity	NTU	Turbidimeter (HACH, 2100P)
TOXFP	μ g/ ℓ	Organic halide analyzer (Dohrmann, DX-2000)
THMFP	μ g/ ℓ	Gas Chromatography (Model HP 5890)
TOC(DOC)	mg/ ℓ	Combustion/non-dispersive infrared gas analysis method (TOC Analyzer, Model TOC-5000, SHIMADZU)
UV-254	cm ⁻¹	UV-Spectrophotometer (UV-1201, SHIMADZU)
Particle counter	-	Particle sizing system (Model 770 Accusizer)

•

4.1 Alum FeCl₃

Fig. 4.1 Fig. 4.2 alum FeCl₃ UV₂₅₄
 TOC . alum FeCl₃가
 TOC FeCl₃ .
 FeCl₃ alum flocc
 . 0.1mM UV₂₅₄ TOC
 , UF 0.05mM
 . alum FeCl₃ 0.05mM
 0.075mM UV₂₅₄ TOC , 0.05mM
 .

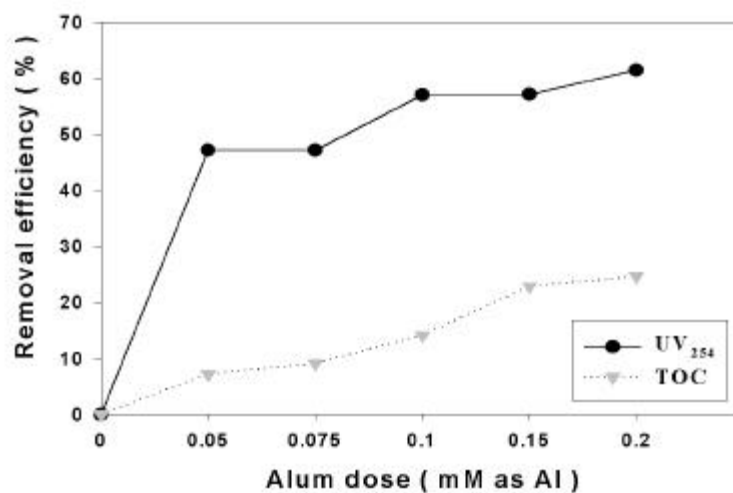


Fig. 4.1 Effect of the sedimentation condition on UV₂₅₄ and TOC removal efficiency with Alum.

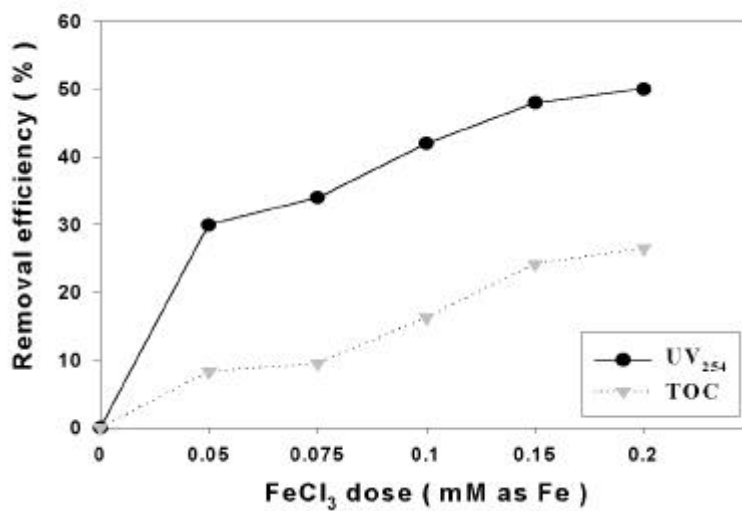


Fig. 4.2 Effect of the sedimentation condition on UV₂₅₄ and TOC removal efficiency with FeCl₃.

4.2

M

4 5

Table 4.1

Table 4.1 Characteristics of raw water

Parameters	Unit	raw water	
Temperature		25	
pH	-	7.8	8.1
Turbidity	NTU	2.1	2.4
UV254	cm ⁻¹	0.061	0.075
TOC	mg/L	3.7	5.21
Alkalinity	mg/L as CaCO ₃	45	55
Conductivity	μ s/cm	194	

Table 4.2 Fig. 4.3 M

DOC

. Fig 4.3

hydrophilic 48.2 %,

fulvic acid 38.8 %, Humic acid 13%

Table 4.2 DOC, UV_{254} and SUVA for organic fraction in raw water

Fraction	DOC (mg/l)	$UV_{254} (cm^{-1})$	SUVA (1/m/ (mg/l))
Humic acid	0.65	0.015	2.31
Hydrophilic	2.411	0.020	0.83
Fulvic acid	1.942	0.037	1.91

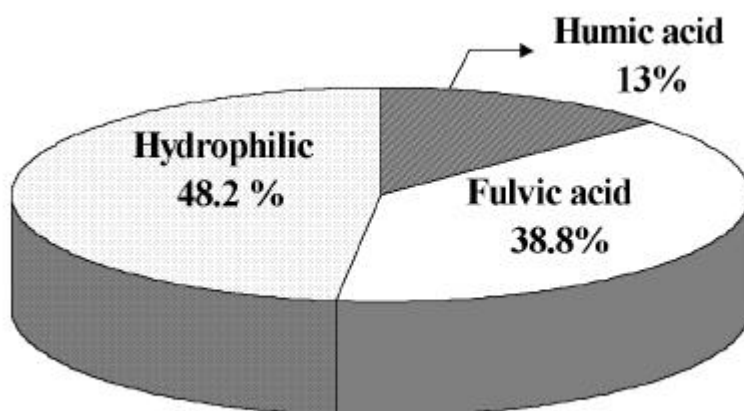


Fig. 4.3 Distribution of organic fraction for DOC.

4.3

THMFP, TOXFP

Fig. 4.4 Fig. 4.5 M

THMFP TOXFP . THMFP TOXFP

TOXFP , hydrophilic 60%, hydrophobic 40%

THMFP hydrophilic 30%, hydrophobic

70% . , hydrophobic

hydrophilic TOX THM

hydrophobic . Rook(1974)

polyhydroxy aromatic 가 humic substance가 THM

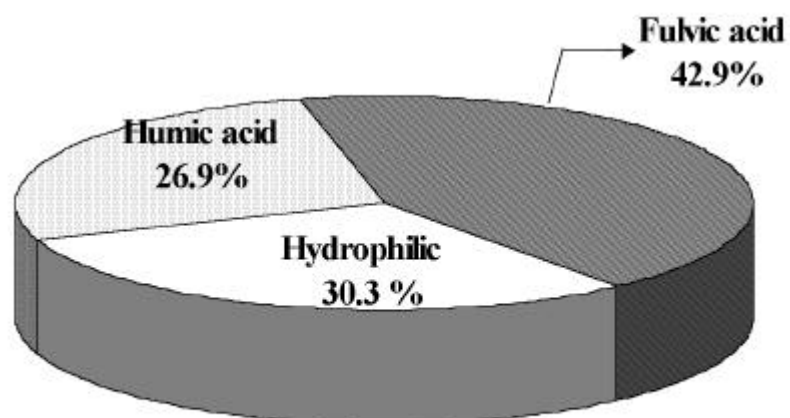


Fig. 4.4 THMFP of DOC fractions for Nakdong river water.

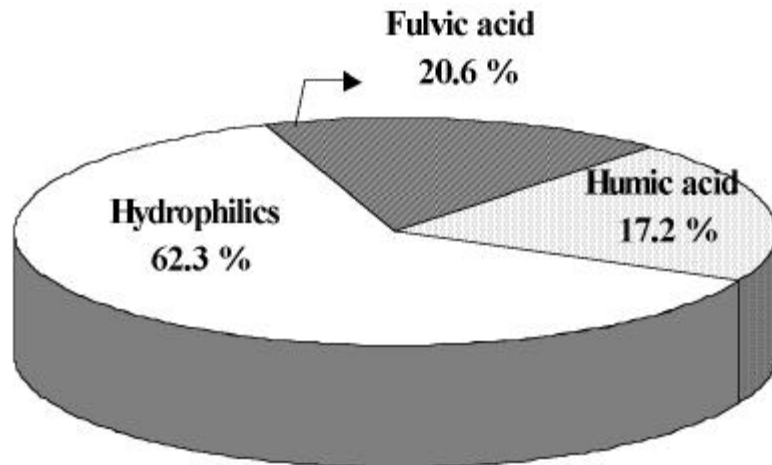


Table 4.3 Present the THMFP and TOXFP of the different extracts on Nakdong river water

Fraction	THMFP (ug/L)	TOXFP (ug/L)	STHMFP* (ug/mg C)	STOXFP** (ug/mg)
hydrophilic	33.3	268.15	13.59	109.45
humic	47.18	73.88	84.86	113.66
fulvic	29.54	88.52	11.9	45.58

*STHMFP : Specific Total Halide Formation Potential (=THMFP/DOC)

**STOXFP : Specific Trihalomethane Formation Potential (=TOXFP/DOC)

Table 4.3 THMFP, TOXFP DOC

specific THMFP, specific THMFP . Table

4.3 humic acid

THMFP 가

, TOXFP Hydrophilic 가

. STHMFP(specific THMFP) STOXFP(specific TOXFP)

humic acid 가 . Legube et al.(1990)

UV₂₅₄ humic acid

fulvic acid가 THMFP TOXFP

. STHMFP STOXFP humic

acid 가

polyhydroxy aromatic 가 humic

.

. Croue Martin(1993) polyhydroxy

aromatic humic acid THMFP TOXFP
 , humic acid
 fulvic acid hydrophilic acids 가 .

4.4 (AMW)

UF Fig. 4.6 .
 MWCO(Molecular Weight Cutoffs) 500, 3,000, 10,000, 30,000(YC05,
 YM3, YM10, YM30) 4 UF .
 500 daltons 47.2% 가
 30,000 daltons 9.1% .

30K UF
 . Lahoussine(1990)
 UF ,

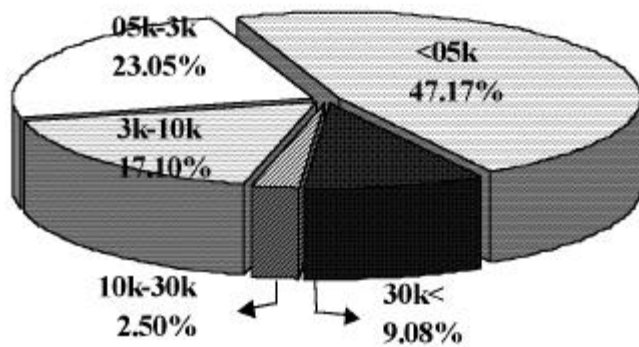


Fig. 4.6 AMW distribution of DOC in raw water for DOC.

Fig. 4.7 Fig. 4.8

	THMFP	THMFP	TOXFP	
		30K Daltons		38.0
%	, 10-30K Daltons	31.9 %		
TOXFP	500 Daltons		36.1%	
TOX				
	THMFP		TOXFP	
				Owen (1993)
30K	THM			

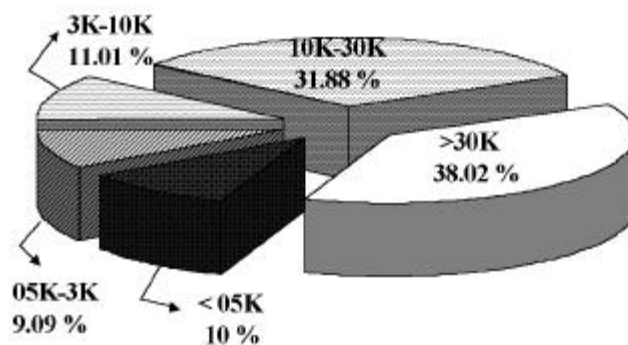


Fig. 4.7 THMFP fraction of DOC according to AMW distribution.

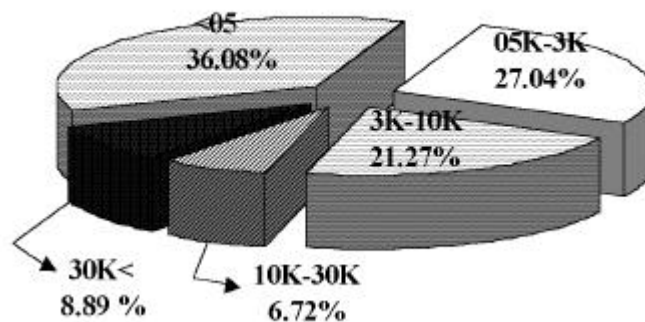


Fig. 4.8 TOXFP fraction for AMW distribution.

Fig. 4.9 Fig. 4.10

STHMFP, STOXFP

THMFP 10K THM

40% , TOX THM

05K

STHMFP STOXFP DOC

10K THM 05K

TOX

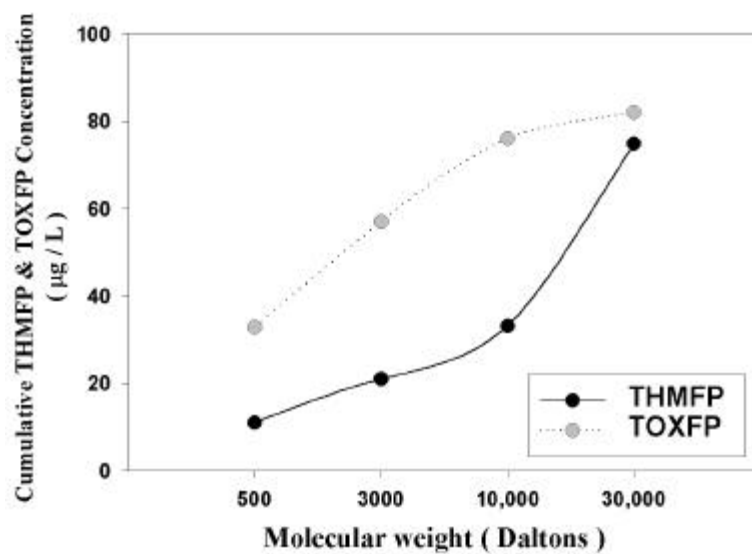


Fig. 4.9 THMFP, TOXFP concentration as a molecular weight accumulate.

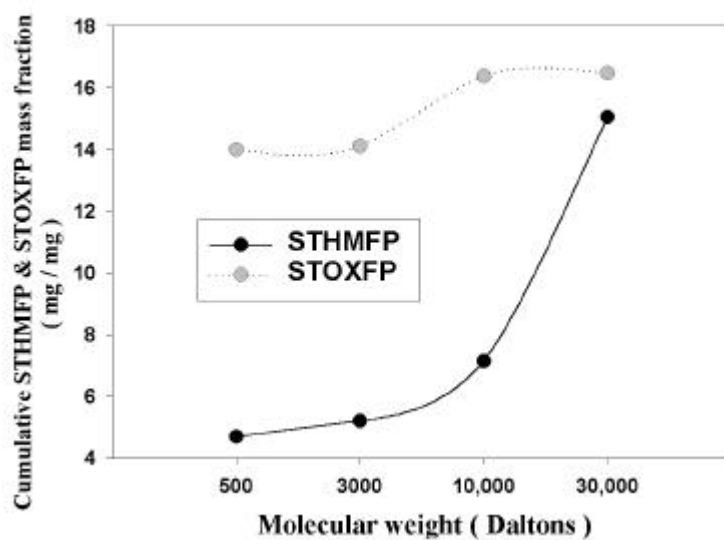


Fig. 4.10 STHMFP, STOXFP mass fraction as a molecular weight accumulate.

4.5 · UF

Fig. 4.11 Alum FeCl_3 humic acid . humic fulvic acid hydrophilic , humic acid fulvic acid , 14% , hydrophilic alum , 7.3% , FeCl_3 11.4% . FeCl_3 , fulvic acid , hydrophilic 12% . FeCl_3 alum (1998) FeCl_3 가 alum .

Fig. 4.12 $\text{Al}(\text{OH})_3$ $\text{Fe}(\text{OH})_3$ UF , . $\text{Al}(\text{OH})_3$ $\text{Fe}(\text{OH})_3$, 가 , humic acid humic . TOX 가 hydrophilic alum FeCl_3 . FeCl_3 UF . (2000) FeCl_3 Alum particle UF 가 .

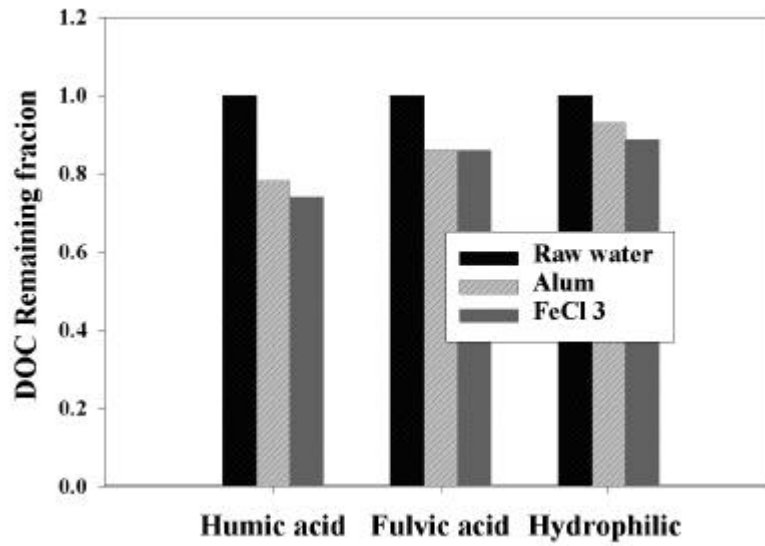


Fig. 4.11 Changes in organic fraction after coagulation process for each coagulant.

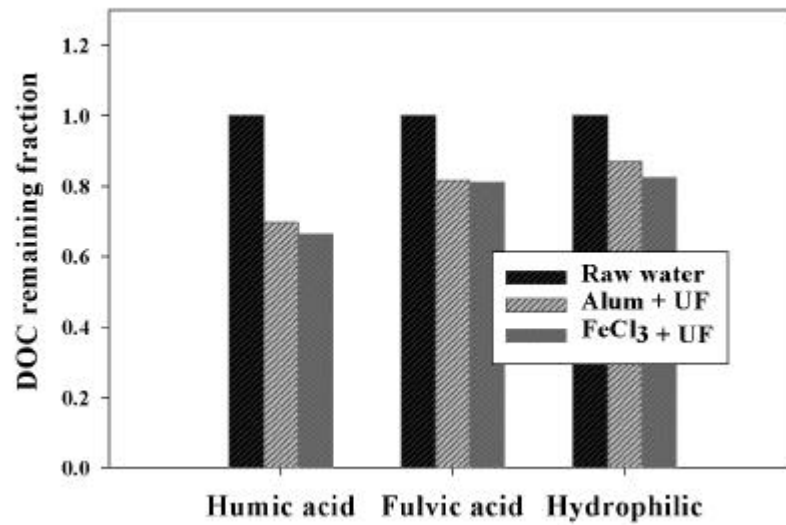


Fig. 4.12 Changes in organic fraction after coagulation and UF process for each coagulant(UF MWCO: 30K).

Table 4.4 , UF
alum 0.05mM(as Al) FeCl₃ 0.05mM(as Fe)
table 4.4
UF
0.05mM
UF
particle
alum FeCl₃
FeCl₃ alum floc
, UF alum

Table. 4.4 Organic matter removal efficiency by coagulation process and coagulation
UF process (Unit: mg/L)

	Humic acid	Fulvic acid	Hydrophilic	Total
Raw water	0.65	1.942	2.411	5.00
Alum	0.504(22.5%)	1.675(13.8%)	2.236(7.3%)	4.415
FeCl ₃	0.481(26.1%)	1.667(14.2%)	2.137(11.4%)	4.285
Alum + UF	0.452(30.5%)	1.582(18.5%)	2.098(13.0%)	4.402
FeCl ₃ + UF	0.431(33.7%)	1.574(19.0%)	1.986(17.6%)	3.991

Fig. 4.13 0.05mM(as Al) , MWCO 30K UF
 , + UF, TOC, UV₂₅₄
 . 30K UF 16% TOC
 .
 30K 가 9.8% UF
 16% TOC .
 TOC 13%, UV₂₅₄ 42% . +UF
 +UF TOC 26%, 28%, UV₂₅₄ 53%, 54% .
 UF UF
 0.05mM(as Al)

Fig. 4.14 0.05mM(as Fe) , MWCO 30K UF
 , +UF, TOC, UV₂₅₄ .
 30K UF 16% TOC
 . 30K
 가 9.8% UF 16%
 TOC . TOC 17%,
 UV₂₅₄ 42% . +UF +UF
 TOC 26%, 32%, UV₂₅₄ 48%, 53% .
 UF TOC

Fig. 4.13

Fig 4.15

THMFP . UF
 UF 15%
 . alum FeCl₃
 FeCl₃ .

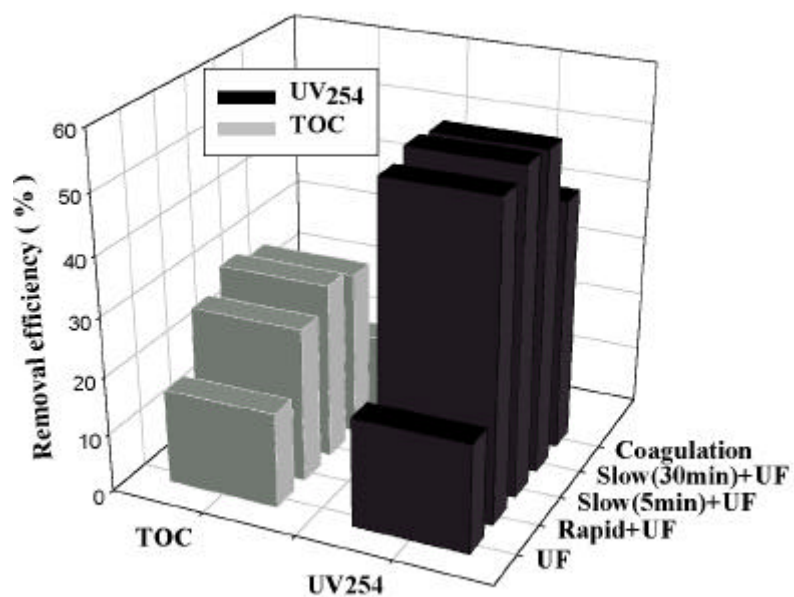


Fig. 4.13 Comparison of UV₂₅₄, TOC removal efficiency after rapid, slow mixing under different mixing time (Coagulant dose : Alum 0.05mM as Al).

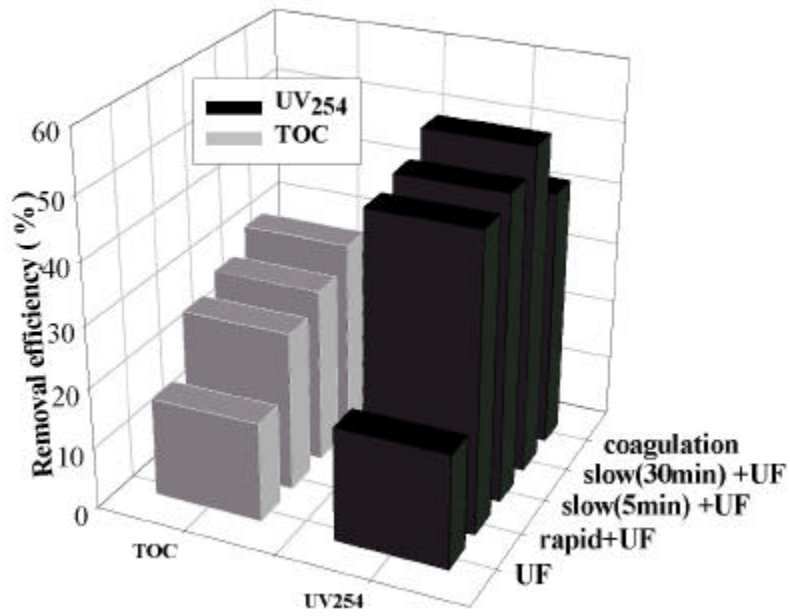


Fig. 4.14 Comparison of UV₂₅₄, TOC removal efficiency after rapid, slow mixing under different mixing time (Coagulant dose: FeCl₃ 0.05mM as Fe).

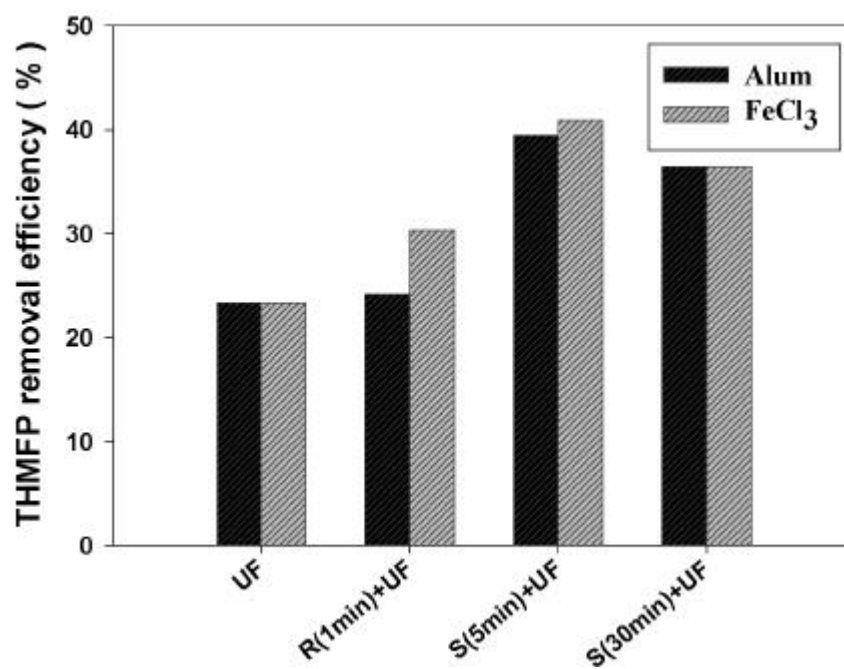


Fig. 4.15 Comparison of THMFP removal efficiency for each process(R+UF: Rapid mixing+UF, S+UF: Slow mixing+UF).

4.6 UF flux

Fig. 4.16 alum UF
flux .
hydrophilic MWCO 30K , 42psi
0.05mM (as Al) . Flux
flux flux flux
(rapid mixing) (slow mixing)
가 .

Fig. 4.16 UF
flux .
flux flux
flux , 5min 30min flux
floc 가 가 .
Kozeny (2-5) ,

$$R = \frac{180(1 - \epsilon_c)^2}{d_p^2 \epsilon_c^3} \text{-----} (2-5)$$

, ϵ_c = cake

d_p = deposit

Cake
floc , flux ,
flux
floc . Laine et al.(1990)
가 UF
flux .

30min flux , flux
 . UF floc
 가
 .
 Fig. 4.17 FeCl₃ , UF
 flux , hydrophilic
 MWCO 30K , 42psi
 0.05mM(as Al) . Fig. 4.17
 UF flux UF
 UF flux 가 UF flux
 . , 30min ,
 UF UF flux UF
 flux . 5min, 30min
 FeCl₃ flux flux .
 alum 5min, 30min 5min
 , FeCl₃
 5min, 30min flux flux
 .

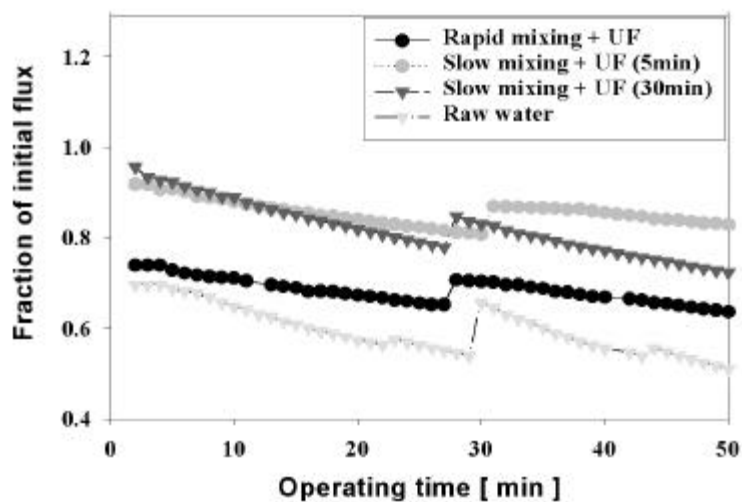


Fig. 4.16 Comparison of flux decline produced by Nakdong river water with and without coagulation pretreatment with alum 0.05mM.

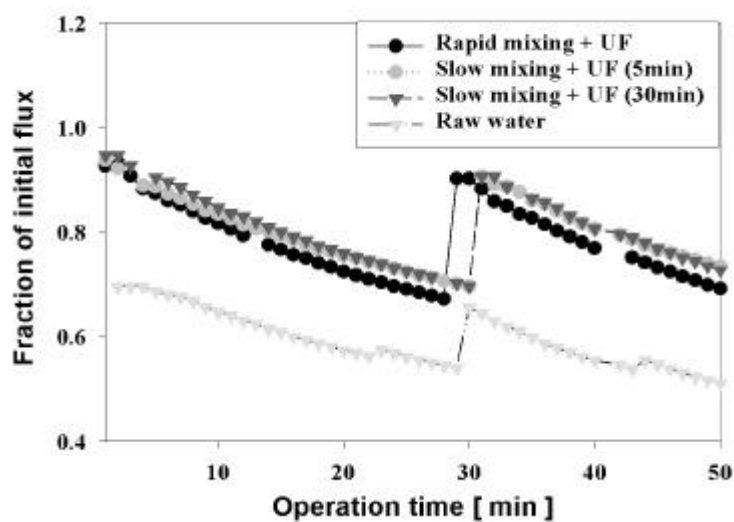


Fig. 4.17 Comparison of flux decline produced by Nakdong river water with and without coagulation pretreatment with FeCl_3 0.05mM.

4.7 UF specific cake resistance

flux UF flux
(α_p)

Fig. 4.18 Fig. 4.19

alum FeCl_3

UF, k Kozeny

(2-5)

가

$J(t)$

$$J(t) = \frac{1}{A_m} \frac{dV}{dt} = \frac{\Delta P}{\mu(R_m' + R_c(t))} \quad (4-1)$$

$$R_m' = R_m + R_b \quad (4-2)$$

$$R_c(t) = \alpha V C_{b,m} t \quad (4-3)$$

$$R_c(t) \quad (4-4) \quad (V C_{b,m} t)$$

, α (4-3) (4-1)

, t/V V α

$$\frac{t}{V} = \frac{R_m' \mu}{\Delta P A_m} + \left(\frac{\alpha C_{b,m} \mu}{2 A_m^2 \Delta P} \right) V \quad (4-4)$$

, R_m' : membrane resistance (m^{-1})

$C_{b,m}$: particle concentration (mg/L)

V : volume (m^3)

t : time (sec)

μ : viscosity (N/sec · m²)

α : specific resistance

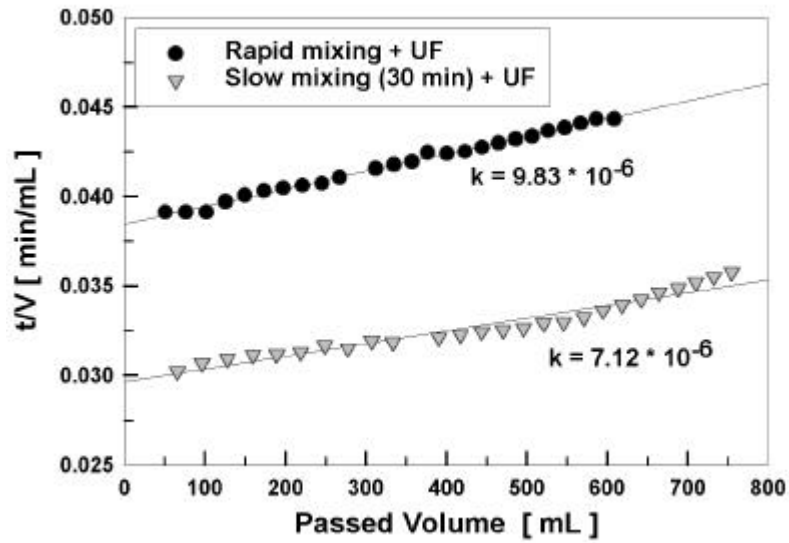


Fig. 4.18 Comparison of typical t/V vs. V curve after rapid and slow mixing (Coagulant: Alum 0.05mM as Al).

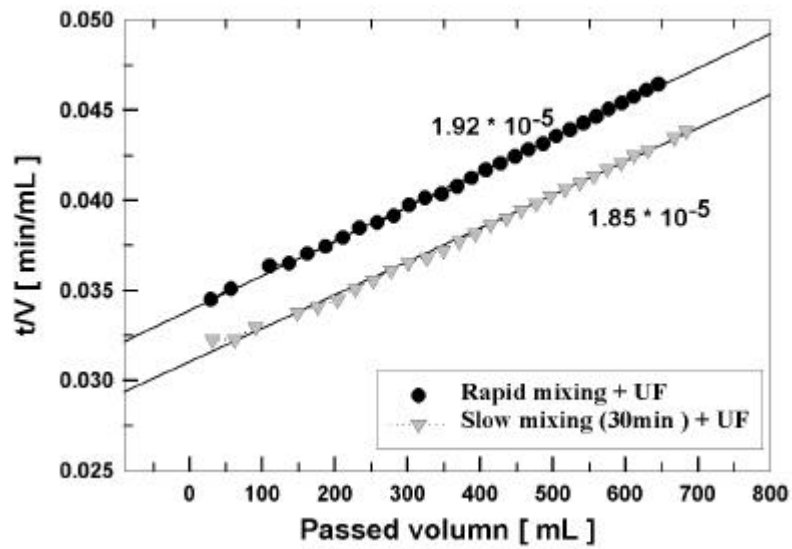
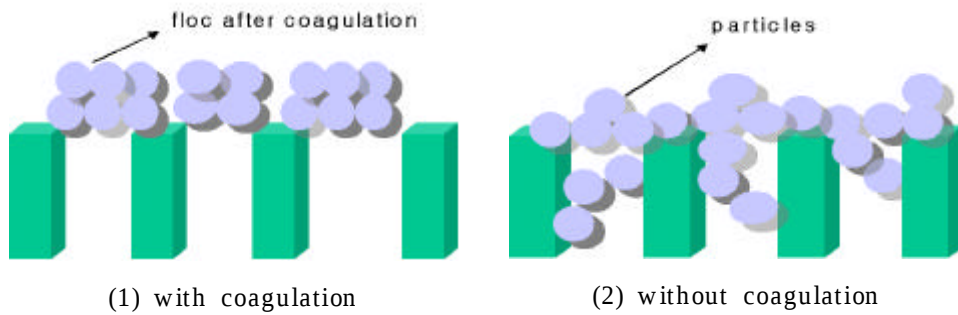


Fig. 4.19 Comparison of typical t/V vs. V curve after rapid and slow mixing (Coagulant: FeCl_3 0.05mM as Fe).

4.8

Fig. 4.20 Fig. 4.21 UF , flux
 , flux , particle size
 1.2 μ m filter
 . Fig. 4.20
 ,
 , 1 μ m
 . Fig. 4.21 Fe()
 , 1 μ m 10 μ m
 10% FeCl₃
 microfloc 가 alum 1 μ m
 가 가 . UF
 가 .



(1) (2) 가
 가 , 가
 floc 가
 cake
 cake
 flux .
 cake
 cake

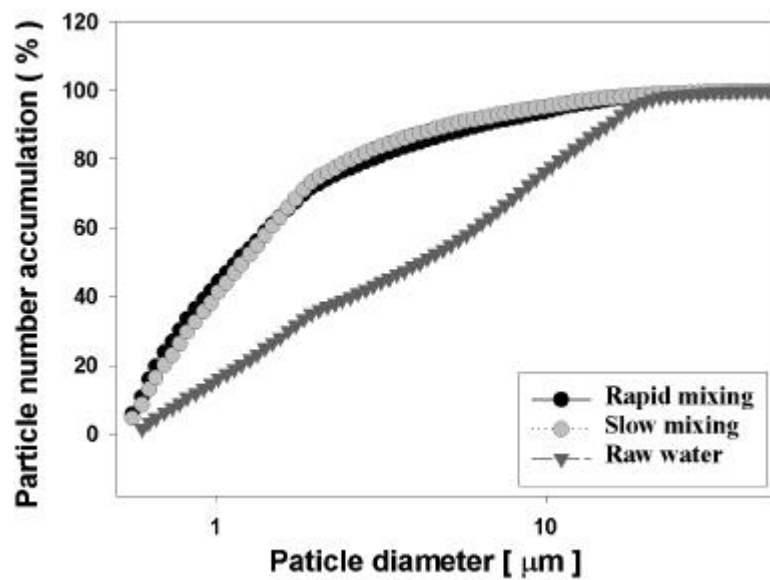


Fig. 4.20 Changes in particle size distribution after rapid, slow mixing (Coagulant: Alum 0.05mM as Al).

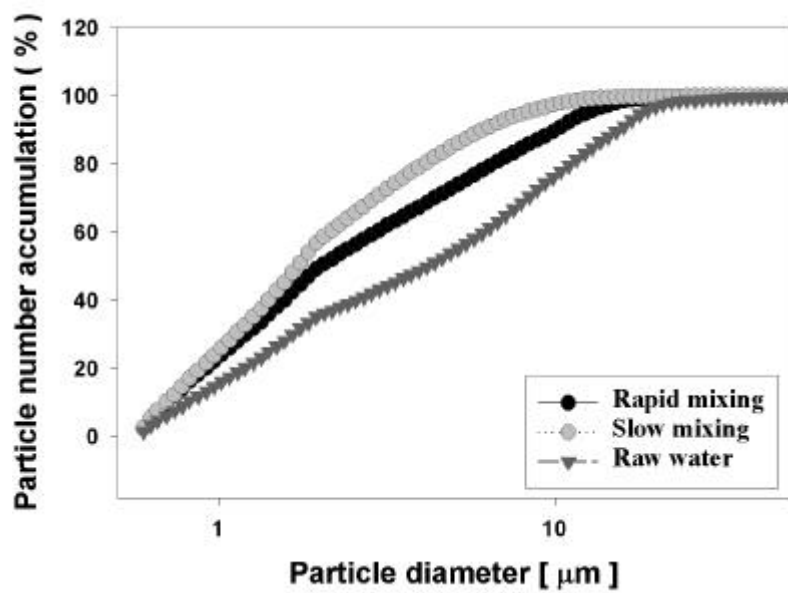


Fig. 4.21 Changes in particle size distribution after rapid, slow mixing (Coagulant: FeCl₃ 0.05mM as Fe).

•

1. , hydrophilic 48.2%, humic acid 13%, fulvic acid 38.8% .

2. THMFP, TOXFP , TOXFP , hydrophilic 62%, hydrophobic 38% , THMFP , hydrophilic 30%, hydrophobic 70% . STHMFP STOXFP , humic acid 가 .

3. , 500daltons 47.2% 가 , 30,000daltons 9.1% . , TOXFP 500daltons 36.1% , THM 30,000daltons 38.1% .

4. , , humic acid . fulvic acid hydrophilic , humic acid hydrophilic alum $FeCl_3$. UF , humic acid . , alum $FeCl_3$ $FeCl_3$ alum hydrophilic .

5. flux UF flux
flux

flux .

6. , alum
FeCl₃ .

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